PATENT ASSIGNMENT COVER SHEET

Electronic Version v1.1 Stylesheet Version v1.2 EPAS ID: PAT5265078

SUBMISSION TYPE:	NEW ASSIGNMENT
NATURE OF CONVEYANCE:	ASSIGNMENT

CONVEYING PARTY DATA

Name	Execution Date
JIM TAYLOR	12/04/2018

RECEIVING PARTY DATA

Name:	SEBASTIAN TAYLOR
Street Address:	1020 E BROAD ST, #112
City:	SAVANNAH
State/Country:	GEORGIA
Postal Code:	31401

PROPERTY NUMBERS Total: 1

Property Type	Number
Patent Number:	8491564

CORRESPONDENCE DATA

Fax Number:

Correspondence will be sent to the e-mail address first; if that is unsuccessful, it will be sent using a fax number, if provided; if that is unsuccessful, it will be sent via US Mail.

Phone: 9176890232

Email: z.sebastiantaylor@gmail.com

Correspondent Name: SEBASTIAN TAYLOR **Address Line 1:** 1020 E BROAD ST

Address Line 2: #112

Address Line 4: SAVANNAH, GEORGIA 31401

NAME OF SUBMITTER:	JIM TAYLOR
SIGNATURE:	/Jim Taylor/
DATE SIGNED:	12/04/2018
	This document serves as an Oath/Declaration (37 CFR 1.63).

Total Attachments: 46

source=UNITED STATES PATENT ADULT MESENCYMAL AUTOLOGOUS STEM CELL (2)#page1.tif source=UNITED STATES PATENT ADULT MESENCYMAL AUTOLOGOUS STEM CELL (2)#page2.tif source=UNITED STATES PATENT ADULT MESENCYMAL AUTOLOGOUS STEM CELL (2)#page3.tif source=UNITED STATES PATENT ADULT MESENCYMAL AUTOLOGOUS STEM CELL (2)#page4.tif source=UNITED STATES PATENT ADULT MESENCYMAL AUTOLOGOUS STEM CELL (2)#page5.tif

PATENT 505218310 REEL: 048383 FRAME: 0272

source=UNITED STATES PATENT ADULT MESENCYMAL AUTOLOGOUS STEM CELL (2)#page6.tif source=UNITED STATES PATENT ADULT MESENCYMAL AUTOLOGOUS STEM CELL (2)#page7.tif source=UNITED STATES PATENT ADULT MESENCYMAL AUTOLOGOUS STEM CELL (2)#page8.tif source=UNITED STATES PATENT ADULT MESENCYMAL AUTOLOGOUS STEM CELL (2)#page9.tif source=UNITED STATES PATENT ADULT MESENCYMAL AUTOLOGOUS STEM CELL (2)#page10.tif source=UNITED STATES PATENT ADULT MESENCYMAL AUTOLOGOUS STEM CELL (2)#page11.tif source=UNITED STATES PATENT ADULT MESENCYMAL AUTOLOGOUS STEM CELL (2)#page12.tif source=UNITED STATES PATENT ADULT MESENCYMAL AUTOLOGOUS STEM CELL (2)#page13.tif source=UNITED STATES PATENT ADULT MESENCYMAL AUTOLOGOUS STEM CELL (2)#page14.tif source=UNITED STATES PATENT ADULT MESENCYMAL AUTOLOGOUS STEM CELL (2)#page15.tif source=UNITED STATES PATENT ADULT MESENCYMAL AUTOLOGOUS STEM CELL (2)#page16.tif source=UNITED STATES PATENT ADULT MESENCYMAL AUTOLOGOUS STEM CELL (2)#page17.tif source=UNITED STATES PATENT ADULT MESENCYMAL AUTOLOGOUS STEM CELL (2)#page18.tif source=UNITED STATES PATENT ADULT MESENCYMAL AUTOLOGOUS STEM CELL (2)#page19.tif source=UNITED STATES PATENT ADULT MESENCYMAL AUTOLOGOUS STEM CELL (2)#page20.tif source=UNITED STATES PATENT ADULT MESENCYMAL AUTOLOGOUS STEM CELL (2)#page21.tif source=UNITED STATES PATENT ADULT MESENCYMAL AUTOLOGOUS STEM CELL (2)#page22.tif source=UNITED STATES PATENT ADULT MESENCYMAL AUTOLOGOUS STEM CELL (2)#page23.tif source=UNITED STATES PATENT ADULT MESENCYMAL AUTOLOGOUS STEM CELL (2)#page24.tif source=UNITED STATES PATENT ADULT MESENCYMAL AUTOLOGOUS STEM CELL (2)#page25.tif source=UNITED STATES PATENT ADULT MESENCYMAL AUTOLOGOUS STEM CELL (2)#page26.tif source=UNITED STATES PATENT ADULT MESENCYMAL AUTOLOGOUS STEM CELL (2)#page27.tif source=UNITED STATES PATENT ADULT MESENCYMAL AUTOLOGOUS STEM CELL (2)#page28.tif source=UNITED STATES PATENT ADULT MESENCYMAL AUTOLOGOUS STEM CELL (2)#page29.tif source=UNITED STATES PATENT ADULT MESENCYMAL AUTOLOGOUS STEM CELL (2)#page30.tif source=UNITED STATES PATENT ADULT MESENCYMAL AUTOLOGOUS STEM CELL (2)#page31.tif source=UNITED STATES PATENT ADULT MESENCYMAL AUTOLOGOUS STEM CELL (2)#page32.tif source=UNITED STATES PATENT ADULT MESENCYMAL AUTOLOGOUS STEM CELL (2)#page33.tif source=UNITED STATES PATENT ADULT MESENCYMAL AUTOLOGOUS STEM CELL (2)#page34.tif source=UNITED STATES PATENT ADULT MESENCYMAL AUTOLOGOUS STEM CELL (2)#page35.tif source=UNITED STATES PATENT ADULT MESENCYMAL AUTOLOGOUS STEM CELL (2)#page36.tif source=UNITED STATES PATENT ADULT MESENCYMAL AUTOLOGOUS STEM CELL (2)#page37.tif source=UNITED STATES PATENT ADULT MESENCYMAL AUTOLOGOUS STEM CELL (2)#page38.tif source=UNITED STATES PATENT ADULT MESENCYMAL AUTOLOGOUS STEM CELL (2)#page39.tif source=UNITED STATES PATENT ADULT MESENCYMAL AUTOLOGOUS STEM CELL (2)#page40.tif source=UNITED STATES PATENT ADULT MESENCYMAL AUTOLOGOUS STEM CELL (2)#page41.tif source=UNITED STATES PATENT ADULT MESENCYMAL AUTOLOGOUS STEM CELL (2)#page42.tif source=UNITED STATES PATENT ADULT MESENCYMAL AUTOLOGOUS STEM CELL (2)#page43.tif source=UNITED STATES PATENT ADULT MESENCYMAL AUTOLOGOUS STEM CELL (2)#page44.tif source=UNITED STATES PATENT ADULT MESENCYMAL AUTOLOGOUS STEM CELL (2)#page45.tif source=Jim Taylor's IP Transfer to Z. Sebastian Taylor#page1.tif

Systems and methods for deriving, preparing and administering adult mesenchymal autologous stem cells

Abstract

The invention provides a system and methodology for deriving, preparing, and administering platelet rich plasma and adult mesenchymal autologous stem cells suitable for regenerating tissue in a living organism. The system includes a sealed primary container capable of removing blood from a living organism. The system further includes a sealed secondary container containing a separation medium and a low-density high-viscosity liquid for receiving the blood. The system further includes a collecting device with a housing and a member wherein the member is moveable in a distal direction is useful in collecting and/or dispensing medium. The system further includes a separation medium capable of separating red blood cells from plasma when the container contains blood and is centrifuged. The system further includes a sealed or unsealed container with a transfer device designed to remove the platelet rich plasma and the adult mesecnhymal autologous stem cells from the blood. The system also includes a transfer device having a apparatus adapted to deliver and/or puncture a fourth container in order to provide fluid communication between the third and fourth containers. The system also includes a transfer device having an apparatus to deliver the adult mesenchymal autologous stem cells to the living organism.

Inventors: Taylor; Jim (Rincon, GA, USA),

Applicant: Name City State Country Type

Taylor: Jim Rincon GA US

Assignee:

Appl. No.: 12/424,317 Filed: April 15, 2009

Related U.S. Patent Documents

Application Number Filing Date Patent Number Issue Date

Foreign Application Priority Data

.....

Jun 24, 1997 [IT] MI97A1490

Current U.S. Class: 604/414

Current International Class: A61B 19/00 (20060101)

Field of Search: 604/403-416,8-10,57 210/650,651,781,782,787

422/101,102

References Cited [Referenced By]

U.S. Patent Documents		
1593814	July 1926	Robert
<u>2006451</u>	July 1935	Glidden
<u>3604410</u>	September 1971	Whitacre
<u>3628974</u>	December 1971	Battista
<u>3654925</u>	April 1972	Holderith
<u>3706305</u>	December 1972	Berger et al.
<u>3883574</u>	May 1975	Axen et al.
<u>3939822</u>	February 1976	Markowitz
<u>3948875</u>	April 1976	Cohen et al.
<u>3981488</u>	September 1976	Ratowsky
4050451	September 1977	Columbus
<u>4091802</u>	May 1978	Columbus
D253190	October 1979	Bixler et al.
<u>4177261</u>	December 1979	Dietze et al.
<u>4251510</u>	February 1981	Tankersley
<u>4272521</u>	June 1981	Zuffi
<u>4272523</u>	June 1981	Kotitschke et al.
<u>4273871</u>	June 1981	Tolbert et al.
<u>4277159</u>	July 1981	Descotes
<u>4277185</u>	July 1981	Thompson
<u>4287180</u>	September 1981	Thomas
<u>4287184</u>	September 1981	Young
<u>4294826</u>	October 1981	Feldman
<u>4296100</u>	October 1981	Franco
4298598	November 1981	Schwarz et al.
<u>4322298</u>	March 1982	Persidsky
4342341	August 1982	Lee
<u>4350687</u>	September 1982	Lipton et al.

4356958	November 1982	Kolobow et al.
4369117	January 1983	White
4378374	March 1983	Reggio et al.
4419089	December 1983	Kolobow et al.
4427650	January 1984	Stroetmann
4427651	January 1984	Stroetmann
4431582	February 1984	Stenn
4444760	April 1984	Thomas
4465669	August 1984	Wissler et al.
4470968	September 1984	Mitra et al.
4470969	September 1984	Pancham et al.
4471053	September 1984	Comi et al.
4479896	October 1984	Antoniades
4479938	October 1984	Thomas
4485096	November 1984	Bell
4503038	March 1985	Banda et al.
4512977	April 1985	Lundy
<u>4514387</u>	April 1985	Wissler
4529590	July 1985	LeVeen et al.
<u>4564359</u>	January 1986	Ruhland
<u>4621052</u>	November 1986	Sugimoto
<u>4639316</u>	January 1987	Eldegheidy
<u>4727137</u>	February 1988	Vallee et al.
<u>4811866</u>	March 1989	Golias
4861477	August 1989	Kimura
4865733	September 1989	Tsuru et al.
<u>4957638</u>	September 1990	Smith
<u>5030215</u>	July 1991	Morse et al.
5037549	August 1991	Ballies
<u>5065768</u>	November 1991	Coleman et al.
<u>5066286</u>	November 1991	Ryan
<u>5163582</u>	November 1992	Godolphin et al.
<u>5165938</u>	November 1992	Knighton
<u>5185001</u>	February 1993	Galanakis
<u>5211310</u>	May 1993	Godolphin et al.
<u>5240856</u>	August 1993	Goffe et al.
<u>5246666</u>	September 1993	Vogler et al.
<u>5257633</u>	November 1993	Vogler et al.

5275731	January 1994	Jahn
5322515	June 1994	Karas et al.
5326535	July 1994	Vogler et al.
<u>5354483</u>	October 1994	Furse
5378431	January 1995	Vogler et al.
<u>5389265</u>	February 1995	Luoma
<u>5413246</u>	May 1995	Godolphin et al.
<u>5419835</u>	May 1995	Adams et al.
<u>5455009</u>	October 1995	Vogler et al.
<u>5456885</u>	October 1995	Coleman et al.
<u>5466065</u>	November 1995	Catrombon
<u>5505683</u>	April 1996	Geringer et al.
<u>5533518</u>	July 1996	Vogler
<u>5555920</u>	September 1996	Godolphin et al.
<u>5560830</u>	October 1996	Coleman et al.
<u>5578459</u>	November 1996	Gordon et al.
<u>5585007</u>	December 1996	Antanavich et al.
<u>5588946</u>	December 1996	Graham et al.
5599718	February 1997	Gorog
<u>5634474</u>	June 1997	Grippi
<u>5634893</u>	June 1997	Rishton
<u>5642938</u>	July 1997	Nakagawa et al.
5643192	July 1997	Hirsh et al.
<u>5667963</u>	September 1997	Smith et al.
<u>5674458</u>	October 1997	Holm
<u>5733545</u>	March 1998	Hood
5736033	April 1998	Coleman et al.
<u>5738670</u>	April 1998	Grippi
5739288	April 1998	Edwardson et al.
<u>5763410</u>	June 1998	Edwardson et al.
<u>5772643</u>	June 1998	Howell et al.
<u>5795489</u>	August 1998	Holm
<u>5795571</u>	August 1998	Cederholm-Williams et al.
<u>5853600</u>	December 1998	McNeal et al.
<u>5935051</u>	August 1999	Bell
<u>5962420</u>	October 1999	Edwardson et al.
<u>6010627</u>	January 2000	Hood
6063297	May 2000	Antanavich et al.

6083383	July 2000	Huang et al.	
6114135	September 2000	Goldstein	
6153104	November 2000	Robertson	
6225123	May 2001	Cohen et al.	
6234948	May 2001	Yavilevich	
6238578	May 2001	Fiehler	
6274090	August 2001	Coelho et al.	
6277060	August 2001	Neumann	
6368298	April 2002	Beretta et al.	
<u>6406671</u>	June 2002	DiCesare et al.	
6544751	April 2003	Brandwein et al.	
6569204	May 2003	Aldecoa	
6596180	July 2003	Baugh et al.	
<u>6596708</u>	July 2003	Petrus	
<u>6811777</u>	November 2004	Mishra	
6905612	June 2005	Dorian et al.	
6979307	December 2005	Beretta et al.	
2004/0071786	April 2004	Beretta et al.	
2006/0074394	April 2006	Beretta et al.	
<u>2006/0128016</u>	June 2006	Tokushima et al.	
2008/0190857	August 2008	Beretta et al.	
2008/0199513	August 2008	Beretta et al.	
2009/0258056	October 2009	Beretta et al.	
2009/0317439	December 2009	Turzi et al.	
2010/0015226	January 2010	Turzi et al.	
2011/0020196	January 2011	Grippi et al.	
	Foreign Patent Documents		
0105014	Apr., 1984		EP
0128849	Dec., 1984		EP
0190018	Aug., 1986		EP
0580094	Jan., 1994		EP
0592242	Apr., 1994		EP
0740155	Oct., 1996		EP
0512612	Feb., 1998		EP
1543846	Aug., 2009		EP
2533438	Mar., 1984		FR
2146335	Apr., 1985		GB
01292410	Feb., 1999		IT

58090513	May., 1983	JР
61200903	Sep., 1986	JP
2311761	Dec., 1990	JP
5099917	Apr., 1993	JP
2504915	May., 1994	JP
8320318	Dec., 1996	JP
9501932	Feb., 1997	JP
10243940	Sep., 1998	JP
10277143	Oct., 1998	JP
2000178201	Jun., 2000	JP
2002022735	Jan., 2002	JP
2003517272	May., 2003	JP
86/05683	Oct., 1986	WO
87/01728	Mar., 1987	WO
91/09573	Jul., 1991	WO
94/22503	Oct., 1994	WO
95/05849	Mar., 1995	WO
95/12371	May., 1995	WO
96/17871	Jun., 1996	WO
96/27397	Sep., 1996	WO
98/11925	Mar., 1998	WO
98/58689	Dec., 1998	WO
00/07659	Feb., 2000	WO
01/60424	Aug., 2001	WO
02/34110	May., 2002	WO
02/45767	Jun., 2002	WO
2006/136870	Dec., 2006	WO
2008/022651	Feb., 2008	WO
2008/023026	Feb., 2008	WO
2011/110948	Sep., 2011	WO

Other References

Office Action from U.S. Appl. No. 10/053,247, dated Nov. 3, 2004 (9 pages). cited by applicant .

Office Action from U.S. Appl. No. 12/032,346, dated Oct. 14, 2008 (8 pages). cited by applicant.

Office Action from U.S. Appl. No. 11/284,584, dated Jun. 1, 2007 (9 pages). cited by applicant.

Office Action from U.S. Appl. No. 11/284,584m dated Sep. 12, 2006 (7pages). cited by applicant.

03796273.5 European Office Action, 4 pages, dated Jun. 23, 2008. cited by applicant.

03796273.5 European Office Action, 3 pages, dated Nov. 30, 2006. cited by applicant.

06027028.7 European Office Action, 6 pages, dated Dec. 23, 2008. cited by applicant.

03703826.2 European Office Action, 6 pages, dated Dec. 12, 2005. cited by applicant.

PCT/US03/01226 International Search Report, 6 pages, dated Sep. 11, 2003. cited by applicant .

PCT/US03/20163 Written Opinion, 4 pages, dated Jun. 22, 2004. cited by applicant . Japanese Patent Application No. 2003-559565 Office Action, 14 pages, dated Nov. 21, 2008. cited by applicant .

Japanese Patent Application No. 2004-557099 Office Action, 13 pages, dated May 1, 2009. cited by applicant.

Office Action from Japanese Patent Office for Application No. 2003-559565 dated Jun. 15, 2009 (12 pages). cited by applicant.

Office Action for U.S. Appl. No. 11/909,191 dated Aug. 5, 2010 (15 pages). cited by applicant.

Office Action for U.S. Appl. No. 12/423,635 dated Nov. 10, 2010 (10 pages). cited by applicant .

Office Action for U.S. Appl. No. 11/909,191 dated Jan. 3, 2011 (14 pages). cited by applicant .

European Patent Office Action for Application No. 10003009.7 dated Jun. 2, 2010 (9 pages). cited by applicant.

Japanese Patent Office Action for Application No. 2004-557099 dated Aug. 30, 2010 (6 pages) with translation. cited by applicant.

Japanese Patent Office Action for Application No. 2009-097789 dated Mar. 28, 2012 (English Translation Only, 2 pages). cited by applicant.

Office Action for U.S. Appl. No. 12/782,617 dated May 18, 2012 (9 pages). cited by applicant .

Annunziata, M. et al., "In vitro cell-type specific biological response of human periodontally related cells to platelet-rich plasma," J. Periodontal Res., 2005, vol. 40, 489-495. cited by applicant.

Castillo, T. et al., "Comparison of Growth Factor and Platelet Concentration From Commercial Platelet-Rich Plasma Separation Systems," AJSM PreView, Nov. 4, 2010. cited by applicant .

Celotti, F. et al., "Effect of platelet-rich plasma on migration and proliferation of SaOS-2 osteoblasts: role of platelet-derived growth factor and transforming growth factor-B," Wound Rep. Reg., 2006, vol. 14, 195-202. cited by applicant.

Currie, L. et al., "The Use of Fibrin Glue in Skin Grafts and Tissue-Engineered Skin Replacements: A Review," Plast. Reconstr. Surg., Nov. 2001, vol. 108, No. 6, 1713-1726. cited by applicant.

Doucet, C. et al., "Platelet Lysates Promote Mesenchymal *Stem Cell* Expansion: A Safety Substitute for Animal Serum in Cell-Based Therapy Applications," J. Cell. Physiol., 2005, vol. 205, 228-236. cited by applicant.

Fulton, James E., "Breast Contouring with `Gelled` Autologous Fat: A 10-Year Update," Intl. J. of Cosmetic Surgery and Aesthetic Dermatology, 2003, vol. 5, No. 2,

155-163. cited by applicant.

Graziani, F. et al., "In vitro effects of different concentration of PRP on primary bone and gingival cell lines. Preliminary results," Minerva Stomatol., Jan.-Feb. 2005, vol. 54, Nos. 1-2, 15-22. cited by applicant.

Green, H. et al., "Growth of cultured human epidermal cells into multiple epithelia suitable for grafting," Proc. Natl. Acad. Sci., Nov. 1979, vol. 76, No. 11, 5665-5668. cited by applicant.

Jung, R. et al., "Platelet-rich plasma and fibrin as delivery systems for recombinant human bone morphogenetic protein-2," Blood Recovery Systems, Inc., Nov. 14, 2005, http://www.bloodrecovery.com/articles.sub.--03.html. cited by applicant.

Kawazoe, T. et al., "Concerning the Possible Application of Medical Treatment Injections with PRP (Platelet Rich Plasma)," The 2007 Research Council Meeting of Japan Society of Plastic and Reconstructive Surgery, 2007, (63 pages--Original and English Translation). cited by applicant.

Leitner, G.C., et al., "Platelet content and growth factor release in platelet-rich plasma: a comparison of four different systems," Vox Sanguinis, 2006, vol. 91, 135-139. cited by applicant.

Lin, S.S. et al., "Controlled release of PRP-derived growth factors promotes osteogenic differentiation of human mesenchymal stm cells," Conf. Proc. IEEE Eng. Med. Biol. Soc., 2006, vol. 1, 4358-61. cited by applicant.

Liu, L. et al., "Corneal Epitheliotrophic Capacity of Three Different Blood-Derived Preparations," Investigative Ophthalmology & Visual Science, Jun. 2006, vol. 47, No. 6, 2438-2444. cited by applicant.

Liu, Y. et al., "Fibroblast proliferation due to exposure to a platelet concentrate in vitro is pH dependent," Wound Rep. Reg., 2002, vol. 10, No. 5, 336-340. cited by applicant. Okuda, K. et al., "Platelet-rich plasma contains high levels of platelet-derived growth factor and transforming growth factor-beta and modulates the proliferation of periodontally related cells in vitro," J. Periodontol., Jun. 2003, vol. 74, No. 6, 849-57. cited by applicant.

Oliva, A. et al., "Ex vivo expansion of bone marrow stromal cells by platelet-rich plasma: a promising strategy in maxillo-facial surgery," Int. J. Immunopathol. Pharmacol., Jul.-Sep. 2005, vol. 18(3 Suppl), 47-53. cited by applicant.

Parkinson, E. et al., "The Epidermis," Chapter 3 of Culture of Epithelial Cells, 2002, Second Edition, 65-94. cited by applicant.

Powell, D.M. et al., "Recovery from deep-plane rhytidectomy following unilateral wound treatment with autologous platelet gel: a pilot study," Arch. Facial Plast. Surg., Oct.-Dec. 2001, vol. 3, No. 4, 245-50. cited by applicant.

Raffoul, W. et al., "Impact of platelets concentrate and keratinocyte suspension on wound healing--a prospective randomized trial," RegenLab, presented at the European Association of Plastic Surgeons meeting on May 30, 2008 (2 pages). cited by applicant

Raffoul, W. et al., "Impact of platelets concentrate and keratinocyte suspension on wound healing--a prospective randomized trial," presented at the European Association of Plastic Surgeons meeting on May 30, 2008 (16 pages). cited by applicant . Regen Lab, "Innovation in Biological Tissue Regeneration," Presentation 2005, 54 pages. cited by applicant .

Regen Lab, "RegenPRP-Kit," available at least as early as Sep. 26, 2004 (18 pages). cited by applicant.

Regen-Kit, Instructions for use, available at least as early as Apr. 26, 2006 (2 pages). cited by applicant .

Rheinwald, J. et al., "Formation of a Keratinizing Epithelium in Culture by a Cloned Cell Line Derived from a Teratoma," Cell, Nov. 1975, vol. 6, 317-330. cited by applicant.

Rheinwald, J. et al., "Serial Cultivation of Strains of Human Epidermal Keratinocytes: the Formation of Keratinizing Colonies from Single Cells," Cell, Nov. 1975, vol. 6, 331-344. cited by applicant.

Ronfard, V. et al., "Use of human keratinocytes cultured on fibrin glue in the treatment of burn wounds," Burns, 1991, vol. 17, No. 3, 181-184. cited by applicant.

Weibrich, G. et al., "Effect of platelet concentration in platelet-rich plasma on perimplant bone regeneration," Bone, 2004, vol. 34, 665-671. cited by applicant.

Weibrich, G. et al., "Growth stimulation of human osteoblast-like cells by thrombocyte concentrates in vitro," Mund. Kiefer Gesichtschir., May 2002, vol. 6, No. 3, 168-74. cited by applicant.

Office Action for U.S. Appl. No. 12/423,635 dated Jun. 17, 2011 (14 pages). cited by applicant.

Office Action for U.S. Appl. No. 12/782,617 dated Dec. 7, 2011 (11 pages). cited by applicant .

European Patent Office Action for Application No. 10003009.7 dated Mar. 16, 2011 (6 pages). cited by applicant.

European Patent Office Action for Application No. 10011683.9 dated Dec. 28, 2010 (5 pages). cited by applicant.

Japanese Patent Office Action for Application No. 2010-006698 dated Aug. 15, 2012 (English Translation Only, 2 pages). cited by applicant.

Canalis E, "Effect of Platelet-Derived Growth Factor on DNA and Protein Synthesis in Cultured Rat Calvaria", Metabolism, vol. 30, No. 10, pp. 970-975, Oct. 1981. cited by applicant.

Niewiarowski S, et al., "Inhibition of the Platelet-Dependent Fibrin Retraction by the Fibrin Stabilizing Factor (FSF, Factor XIII)", The Journal of Laboratory and Clinical Medicine, vol. 81, No. 5, pp. 641-650, May 1973. cited by applicant.

Li Akc, et al., "Mechanical and Humoral Factors in Wound Healing", British Journal of Surgery, vol. 68, pp. 738-743, 1981. cited by applicant.

Niewiarowski S, et al., "Potentiation of the Thrombin Induced Platelet Release Reaction by Fibrin", Thrombosis Research, vol. 9, No. 2, pp. 181-190, 1976. cited by applicant .

Carroll RC, et al., "Clot Retraction Facilitates Clot Lysis", Blood, vol. 57, No. 1, pp. 44-48, Jan. 1981. cited by applicant .

Joist JH, et al., "Retention of Platelet Fibrin Stabilizing Factor During the Platelet Release Reaction and Clot Retraction", Thrombos. Diathes. Haemorrh., vol. 29, pp. 679-683, 1973. cited by applicant.

Niall M, et al., "The Effect of Epidermal Growth Factor on Wound Healing in Mice", Journal of Surgical Research, vol. 33, No. 2, pp. 164-169, Aug. 1982. cited by applicant.

Cazenave JP, et al., "Inhibition of Platelet Adherence to a Collagen-Coated Surface by Agents that Inhibit Platelet Shape Change and Clot Retraction", Journal of Laboratory and Clinical Medicine, vol. 84, No. 4, pp. 483-493, Oct. 1974. cited by applicant. Niewiarowski S, et al., "Effect of ADP and Thrombin on Fibrin Retraction Induced by Human Platelets and Fibroblasts", Thromb Diathes Haemorrh, vol. 34, pp. 316-317, Oct. 1974. cited by applicant.

Niewiarowski S, et al., "Fibrin Clot Retraction by Human Skin Fibroblasts: Effects of ADP and Thrombin", Proceedings of the Society for Experimental Biology and Medicine, vol. 151, pp. 253-256, 1976. cited by applicant.

Cohen I, et al., "Fibrin-Blood Platelet Interaction in a Contracting Clot", Thromb Diath Haemorrh, vol. 34, No. 2, p. 559, Nov. 15, 1975. cited by applicant.

Day HJ, et al., "Platelet Release Reaction During Clotting of Native Human Platelet-Rich Plasma", Proceedings of the Society of Experimental and Biological Medicine, vol. 139, No. 3, pp. 717-721, 1972. cited by applicant.

Niewiarowski S, et al., "ADP, Thrombin and Bothrops Atrox Thrombinlike Enzyme in Platelet-Dependent Fibrin Retraction", American Journal of Physiology, vol. 229, No. 3, pp. 737-745, Sep. 1975. cited by applicant.

Mustard JF, et al., "Preparation of Suspensions of Washed Platelets from Humans", British Journal of Haematology, vol. 22, No. 2, pp. 193-204, Feb. 1972. cited by applicant.

Packham MA, et al., "Platelet Aggregation and Release: Effects of Low Concentrations of Thrombin or Collagen", American Journal of Physiology, vol. 225, No. 1, pp. 38-47, Jul. 1973. cited by applicant.

Niewiarowski S, et al., "Platelet Aggregation by ADP and Thrombin", Nature, vol. 212, No. 5070, pp. 1544-1547, Dec. 31, 1966. cited by applicant.

Thorton JW, et al., "Epidermal Growth Factor in the Healing of Second Degree Burns: A Controlled Animal Study", Burns, vol. 8, No. 3, pp. 156-160, Feb. 17, 1981. cited by applicant.

Holmsen H, et al., "The Blood Platelet Release Reaction", Scandinavian Journal of Haematology (Supplement K), pp. 3-21, 1969. cited by applicant.

Rosenthal AR, et al., "Use of a Platelet-Fibrinogen-Thrombin Mixture as a Corneal Adhesive: Experiments in Sutureless Lamellar Keratoplasty in the Rabbit", Investigative Ophthalmology, vol. 14, No. 11, pp. 872-875, Nov. 1975. cited by applicant.

Schulte W, "Centrifuged Autologous Blood for Filling Large Bone Defects: A Modification to the Autologous Blood Method", Zentrifugiertes Eigenblut [Centrifuged Autologous Blood] DZZ 24, vol. 10 pp. 854-857, 1969. cited by applicant. Schutle W, "Autologous Blood Filling: A New Method in the Treatment of Major Bone Defects Following Oral Surgery". Eigenblutfullung groBerer Knochendefekte [Autologous Blood Filling of Major Bone Defects] DZZ 15, vol. 12, pp. 910-914, 1960. cited by applicant.

Rosenthal AR, et al., "Use of Platelet-Fibrinogen-Thrombin Mixture to Seal Experimental Penetrating Corneal Wounds", Graefes Archiv Ophthalmologie, vol. 207, pp. 111-115, 1978. cited by applicant.

Silverberg GD, et al., "A Physiological Sealant for Cerebrospinal Fluid Leaks", Journal of Neurosurgery, vol. 46, pp. 215-219, Feb. 1977. cited by applicant.

Pearl RM, et al., "Microvascular Anastomosis Using a Blood Product Sealant-Adhesive", Surgery, Gynecology & Obstetrics, vol. 144, pp. 227-231, Feb. 1977. cited by applicant.

Wolf G, "Der Konzentrierte Autologe Gewebekleber", Arch Otorhinolaryngol, vol. 237: pp. 279-283, Spring 1983. cited by applicant.

Malkin, A.Y., Rheology Fundamentals, pp. 95, 104, and 245, ChemTec Publishing: Ontario, Canada, Available at:

 1994.cited by applicant">http://www.knovel.com/knovel2/Toc.jsp?BookID=322&VerticalID=0> 1994.cited by applicant .

Office Action from U.S. Appl. No. 10/607,580 dated Sep. 11, 2006 (10 pages). cited by applicant .

Office Action from U.S. Appl. No. 10/607,580 dated Jun. 28, 2007 (9 pages). cited by applicant .

Office Action from U.S. Appl. No. 10/607,580 dated Mar. 21, 2008 (8 pages). cited by applicant.

Office Action from U.S. Appl. No. 10/607,580 dated Oct. 28, 2008 (10 pages). cited by applicant .

Bernstein, Lori R, et al., "Migration of Cultured Vascular Cells in Response to Plasma and Platelet-Derived Factors", Departments of Physiology and Surgery, Harvard Medical School, Children's Hospital Medical Center, J. Cell Sci. 56, pp. 71-82, 1982. cited by applicant.

Zetter, Bruce, R., et al., "Stimulation of Human Vascular Endothelial Cell Growth by a Platelet Derived Growth Factor and Thrombin", Journal of Supramolecular Structure 11, pp. 361-370. cited by applicant.

Knighton, David R., et al., "Platelet-Derived Angiogenesis: Initiator of Healing Sequence", Department of Surgery, School of Medicine, University of California, San Francisco, pp. 226-228. cited by applicant.

Seppa, Heikki, et al., "Platelet-Derived Growth Factor is Chemotactic for Fibroblasts", The Journal of Cell Biology, vol. 92, pp. 584-588, Feb. 1982. cited by applicant . Senior, Robert M, et al., "Chemotactic Activity of Platelet Alpha Granule Proteins for

Fibroblasts", The Journal of Cell Biology, vol. 96, pp. 382-385, Feb. 1983. cited by applicant.

Gospodarowicz, Denis, "Humoral Control of Cell Proliferation: The Role of Fibroblast Growth Factor in Regeneration, Angiogenesis, Wound Healing, and Neoplastic Growth", Department of Molecular Endocrinology, Salk Institute for Biological Studies, San Diego, California, pp. 1-19. cited by applicant.

Franklin JD, et al., "Effects of Topical Application of Epidermal Growth Factor on Wound Healing", Plastic and Reconstructive Surgery, vol. 64, No. 6, pp. 766-770, Dec. 1979. cited by applicant.

Buckley A, et al., "Sustained Release of Epidermal Growth Factor Accelerates Wound Repair", Proceeding National Academy Science USA, vol. 82, pp. 7340-7344, Nov. 1985. cited by applicant.

Sporn MB, et al., "Polypeptide Transforming Growth Factors Isolated from Bovine Sources and Used for Wound Healing in Vivo", Science, vol. 219, pp. 1329-1331-, Mar. 18, 1983. cited by applicant.

Dhall TZ, et al., "Fibrin Network Structure: Modification by Platelets", Thromb

Haemostas vol. 49, No. 1, pp. 42-46, 1983. cited by applicant.

Niewiarowski S, et al., "Platelet Interaction with Fibrinogen and Fibrin: Comparison of the Interaction of Platelets with that of Fibroblasts, Leukocytes and Erythrocytes", Annals New York Academy of Sciences, pp. 72-83. cited by applicant.

Knighton DR, et al., "Role of Platelets and Fibrin in the Healing Sequence: An in Vivo Study of Angiogenesis and Collagen Synthesis", Annals of Surgery, vol. 196, No. 4, pp. 379-388, Oct. 1982. cited by applicant.

Doni MG, et al., "Thrombin-Induced Calcium and Magnesium Platelet Release and Clot Retraction", Haematologica, vol. 60, No. 3, pp. 286-299, Sep. 1975. cited by applicant.

Jelenska M, et al., "Blood Platelets Cause Retraction of Collagen Gel", Department of Radiobiology and Health Protection, Institute of Nuclear Research, Warsaw, Poland, pp. 161-164, 1980. cited by applicant.

Biggs, R., Table of Contents, Journal of the International Society on Thrombosis & Haemostasis, Thrombosis et Diathesis Haemorrhagica, pp. III-VI. cited by applicant. Lundblad RL, "A Rapid Method for the Purification of Bovine Thrombin and the Inhibition of the Purified Enzyme with Phenylmethylsulfonyl Fluoride", Biochemistry, vol. 10, No. 13, pp. 2501-2505, Jun. 22, 1971. cited by applicant.

Chao FC, et al., "Concentration Effects of Platelets, Fibrinogen, and Thrombos of Platelet Aggregation and Fibrin Clotting", Thrombos. Diathes. Haemorrh., vol. 32, pp. 216-231, 1974. cited by applicant.

Solum NO, "Platelet Aggregation During Fibrin Polymerization", Scandinavian Journal of Clinical & Laboratory Investigation, vol. 18, No. 216, pp. 577-587, 1966. cited by applicant.

Tashjian, "Platelet-Derived Growth Factor Stimulates Bone Resorption Via a Prostaglandin-Mediated Mechanism", Endocrinology, vol. 111, No. 1, pp. 118-124, 1982. cited by applicant.

Mustard JF, et al., "Factors Responsible for ADP-Induced Release Reaction of Human Platelets", American Journal of Physiology, vol. 228, No. 6, Jun. 1975. cited by applicant.

Primary Examiner: Wiest; Philip R

Attorney, Agent or Firm: Michael Best & Friedrich LLP

Claims

We claim:

- 1.A system for deriving, preparing and administering platelet rich plasma and adult mesenchymal autologous stem cells suitable for regenerating tissue in a living organism; the system comprising a sealed primary container having a single opening configured and capable of removing blood from a living organism; the system further including a sealed secondary container containing a separation medium mixed with the receiving blood. The system further includes a collecting device with a housing and a member wherein the member is moveable in a distal direction and is useful in collecting and/or dispensing medium. The system further includes a separation medium capable of separating red blood cells from platelet rich plasma and adult mesenchymal autologous stem cells when the container contains blood and is centrifuged; a sealed or unsealed tertiary container with a transfer device designed to remove the platelet rich plasma and the adult mesenchymal autologous stem cells from the blood; a transfer device having a apparatus adapted to deliver and/or puncture a fourth container in order to provide fluid communication between the third and fourth containers. The system also includes a transfer device having an apparatus to deliver the platelet rich plasma and adult mesenchymal autologous stem cells to the living organism.
- 2. The system of claim 1, wherein the primary container removes blood from the living organism and is delivered into a secondary container.
- 3. The system of claim 1, wherein the secondary container contains a separation medium of ionic coagulation activator is selected from but not limited to; heparin, bivalirudin, a similar derivative and/or combinations thereof.
- 4. The system of claim 1, wherein the system includes a collecting device with a housing and a member wherein the member is moveable in a distal direction and is useful in collecting and/or dispensing medium.
- 5. The system of claim 1, wherein a separation medium in the secondary container is capable of separating red blood cells from platelet rich plasma and adult mesenchymal stem cells when the container contains blood and is centrifuged.
- 6. The system of claim 1, wherein a sealed and/or unsealed container with a transfer device designed to remove the platelet rich plasma and the adult mesenchymal autologous stem cells from the secondary container leaving only the blood.
- 7. The system of claim 1, wherein a transfer device having an apparatus adapted to deliver and/or puncture a fourth container in order to provide fluid communication between the third and fourth containers.

- 8. The system of claim 1, wherein a transfer device having an apparatus to deliver the platelet rich plasma and adult mesenchymal autologous stem cells to the living organism.
- 9. A system for deriving, preparing and administering platelet rich plasma and adult mesenchymal autologous stem cells for regenerating tissue in a living organism. The system comprising: a primary container capable of removing blood from a living organism, a sealed secondary container containing a separation medium mixed with the receiving blood, a collecting device with a housing and a member wherein the member is moveable in a distal direction is useful in collecting and/or dispensing medium, a separation medium capable of separating red blood cells from platelet rich plasma and adult mesenchymal stem cells when the container contains blood and is centrifuged, a sealed or unsealed container with a transfer device designed to remove the platelet rich plasma and the adult mesenchymal autologous stem cells from the secondary container leaving only the blood, a transfer device having a apparatus adapted to deliver and/or puncture a fourth container in order to provide fluid communication between the third and fourth containers, and a transfer device having an apparatus to deliver the platelet rich plasma and adult mesenchymal autologous stem cells to the living organism.
- 10. The system of claim 9, wherein the primary container is sealed and capable of removing blood from a living organism.
- 11. The system of claim 9, wherein the separation medium of ionic coagulation activator is selected from, but not limited to; heparin, bivalirudin, a similar derivative and/or combinations thereof.
- 12. The system of claim 9, wherein the secondary container further contains an anti-coagulant.
- 13. The system of claim 9, wherein the system includes a collecting device with a housing and a member wherein the member is moveable in a distal direction is useful in collecting and/or dispensing medium.
- 14. The system of claim 9, wherein a separation medium capable of separating red blood cells from platelet rich plasma and adult mesenchymal autologous stem cells when the container contains blood and is centrifuged.
- 15. The system of claim 9, wherein a sealed and/or unsealed container with a transfer device designed to remove the platelet rich plasma and the adult mesenchymal autologous stem cells from the blood.
- 16. The system of claim 9, wherein a transfer device having an apparatus adapted to deliver and/or puncture a fourth container in order to provide fluid communication between the third and fourth containers.
- 17. The system of claim 9, wherein a transfer device having an apparatus to deliver the platelet rich plasma and adult mesenchymal autologous stem cells to the living organism.

- 18. A system for deriving, preparing and administering platelet rich plasma and adult mesenchymal autologous stem cells suitable for regenerating tissue in a living organism, the system comprising: a primary container having a single opening configured to remove blood from a living organism; a secondary container capable of receiving blood and is capable to receive a cap to form a seal therewith, the cap capable of being removed from the container and/or pierced in sealing the container for delivery of blood into the secondary container, the secondary container containing a separation medium capable of separating red blood cells from platelet rich plasma and adult mesenchymal stem cells when the container is centrifuged; a collecting device with a housing and a member wherein the member is moveable in a distal direction is useful in collecting and/or dispensing medium, a sealed or unsealed container with a transfer device designed to remove the platelet rich plasma and the adult mesenchymal autologous stem cells from the blood, the device configured to pierce the cap or in lieu of removing the cap, transfer a portion of the platelet rich plasma and adult mesenchymal autologous stem cells to a device; the device configured to pierce the cap and transfer a portion of the platelet rich plasma and the adult mesenchymal autologous stem cells from the secondary container to the third container via pressure differentiation upon establishment of fluid communication between the cannula of the third container undergoing removal of the platelet rich plasma and the adult mesenchymal autologous stem cells until only the portion of the red blood cells remains in the secondary container.
- 19. The system of claim 18, wherein the device comprises a cannula attached to a collecting device with a housing and a member wherein the member is moveable in a distal direction is useful in collecting and/or dispensing medium, a separation device capable of separating platlet rich plasma and adult mesenchymal autologous stem cells from red blood cells after the container has been centrifuged to separate the platelet rich plasma and the adult mesenchymal autologous stem cells until only the portion of the red blood cells remain in the secondary container.
- 20. The system of claim 18, wherein a transfer device having a apparatus adapted to deliver and/or puncture a fourth container in order to provide fluid communication between the third and fourth containers.
- 21. The system of claim 18, wherein the third container further includes a transfer device having an apparatus to deliver the platelet rich plasma and adult mesenchymal autologous stem cells to the living organism.

Description

This patent application fully incorporates by reference the subject matter of each of the above-identified patent applications to which this application claims priority. The entire disclosure of each patent application is considered to be part of the accompanying application.

BACKGROUND OF THE INVENTION

The present invention relates to a system and methodology for deriving, preparing and administering platlet rich plasma and adult mesenchymal autologous stem cells suitable for regenerating tissue in a living organism.

Adult mesenchymal autologous stem cells are known to divide or self-renew indefinitely and their differentiation potential is unknown and unlimited. Adult mesenchymal autologous stem cell treatments have been successfully used for many years to treat leukemia and related bone/blood cancers through bone marrow transplants.

Several kits are available on the market that contain adult mesenchymal autologous stem cells from donors.

Such known kits involve the use of material of human or animal origin, which, owing to its origin, could result in possible viral contamination and in serious risks for the receiver of the donored adult mesenchymal autologous stem cells. In the past the authorities have been compelled to suspend from trade or even ban the stem cell derivatives obtained by using material of human or animal origin. Furthermore, rejection cases are known from the literature resulting from reimplanting donored adult mesenchymal autologous stem cells produced by using human or animal proteins in patients. Such cases are indeed due to the donored origin, with respect to the receiver organism, of the donored protein being reimplanted or some of the components used for preparing it.

The adult mesenchymal autologous stem cells obtained from a patient's own blood, is more reliable with respect to the rejection and/or infection risks. Several procedures have already been described for obtaining extemporary adult mesenchymal autologous stem cells, but no system and methodology for deriving, preparing and administering adult mesenchymal autologous stem cells suitable for regenerating tissue in a living organism is available on the market although some relevant references with similarities can be found in the patent literature.

U.S. Pat. No. 5,733,545 discloses a plasma-buffy coat concentrate to be combined with a fibrinogen activator to form a platelet glue wound sealant. The method disclosed in this patent allows for a patient's blood to be processed in order to obtain autologous fibrin glue, but the methods use thrombin or batroxobin as the fibrinogen activator. These activators are of human or animal nature and therefore still involve the risk of rejection and/or viral infections for the patient.

U.S. Pat. No. 5,555,007 discloses a method and an apparatus for making concentrated plasma to be used as a tissue sealant. The method consists in separating plasma from whole blood and removing water from said plasma by contacting it with a concentrator to provide concentrated

plasma which can be thereafter coagulated with a solution containing thrombin and calcium. The apparatus comprises a first centrifuge separator in a first chamber, a concentrator (e.g. dextranomer or polyacrylamide) included in a second chamber communicating with the first chamber, and a second separator. The method disclosed in this reference requires a long time for obtaining the plasma concentrate necessary for the subsequent preparation of autologous fibrin glue and the apparatus is expensive and not disposable. The method does not disclose using a calcium-coagulation activator, and requires a pre-concentration step.

Overall, methods and systems for deriving, preparing and administering adult mesenchymal autologous stem cells suitable for regenerating tissue in a living organism are desired.

DETAILED DESCRIPTION OF THE DRAWINGS

- FIG. 1 is a perspective view of a first embodiment of the invention.
- FIG. 2 is a cross-sectional view of a closed container of the first embodiment shown in FIG. 1 wherein the primary container removes blood from the living organism.
- FIG. 3 is a cross-sectional view of a secondary container of the first embodiment shown in FIG. 1 wherein the secondary container contains an anti-coagulant and is capable of receiving blood through the closed container of the first embodiment.
- FIG. 4 is a cross-sectional view of a second embodiment of the invention showing collecting device with a housing and a member wherein the member is moveable in a distal direction is useful in collecting and/or dispensing medium.
- FIG. 5 is a cross-sectional view of a different embodiment of the closed container of the first embodiment shown in FIG. 1 wherein the primary container removes blood from the living organism and is transferred to a secondary container containing an anti-coagulant.
- FIG. 6 is a cross-sectional view of the secondary container of the first embodiment of FIG. 3 housed in a container with a separation medium capable of separating red blood cells from plasma when the container contains blood before being centrifuged.
- FIG. 7 is the cross-sectional view of a device capable of separating red blood cells from plasma when the container contains blood and is centrifuged.
- FIG. 8 is a cross-sectional view of a different embodiment when the secondary container containing blood and a separation medium is placed in a device capable of separating red blood cells from plasma when the container is centrifuged.

- FIG. 9 is a cross-sectional view of a different embodiment after the secondary container containing blood and a separation medium has been placed in a device capable of separating red blood cells from plasma when the container is centrifuged depicting the separation of whole blood, adult mesenchymal autologous stem cells and platelet rich plasma.
- FIG. 10 is a cross-sectional view of a second embodiment of the invention wherein a sealed or unsealed container with a transfer device designed to remove the platelet rich plasma and the adult mesenchymal autologous stem cells from the blood, the device configured to pierce the cap or in lieu of removing the cap is engaged.
- FIG. 11 is a cross-sectional view of FIG. 10 showing the secondary container and transfer device engaged, and the contents of the second container being transferred to the third container of the transfer device leaving only the whole blood in the secondary container.
- FIG. 12 is a cross-sectional view of a different embodiment of the transfer device containing the platelet rich plasma and the adult mesenchymal autologous stem cells.
- FIG. 13 is a cross-sectional view of the third embodiment of the invention wherein the collecting device with a housing and a member wherein the member is moveable in a distal direction, with its contents, is able to dispense its medium into a second medium housed in a container for multiplying, regenerating, duplicating and incubating adult mesencymal autologous stem cells for future use.
- FIG. 14 is a cross-sectional view of the fourth embodiment of the invention wherein the collecting device with a housing and a member wherein the member is moveable in a distal direction, with its contents, is able to dispense its medium into an area that includes but is not limited to; musculoskeletal, ligament, tendon, cartilage, meniscus, and disc of a living organism for regeneration of said area.
- FIG. 15 is a cross-sectional view of the fifth embodiment of the invention wherein the collecting device with a housing and a member wherein the member is moveable in a distal direction, with its contents, is able to dispense its medium into an area that includes but is not limited to; musculoskeletal, ligament, tendon, cartilage, meniscus, and disc of a living organism for regeneration of said area.
- FIG. 16 is a cross-sectional view of the sixth embodiment of the invention wherein the collecting device with a housing and a member wherein the member is moveable in a distal direction, with its contents, is able to dispense its medium into a fifth container that contains but is not limited to sodium chloride or similar solution.
- FIG. 17 is a cross-sectional view of the sixth embodiment of the invention wherein the methodology of dispensing the medium containing the platelet rich plasma and adult mesencymal autologous stem cells into a living organism via intravenous or intra-arterial.

FIG. 18 is a cross-sectional view of the seventh embodiment of the invention wherein the collecting device with a housing and a member wherein the member is moveable in a distal direction, with its contents, is able to dispense its medium into a sixth container that contains but is not limited to albuterol, sodium chloride or similar solution.

FIG. 19 is a cross-sectional view of the seventh embodiment of the invention wherein the methodology of dispensing the medium containing the adult mesencymal autologous stem cells into a living organism via inhalation.

FIG. 20 is a cross-sectional view of a ready-to-use step-by-step system and methodology packaged in such a way that the accoutrements include all of the components required for deriving, preparing and administering adult mesenchymal autologous stem cells suitable for regenerating tissue in a living organism.

Before one embodiment of the invention is explained in detail, it is to be understood that the invention is not limited in its application to the details of construction and the arrangement of components set forth in the following description or illustrated in the drawings. The invention is capable of other embodiments and of being practiced or being carried out in various ways. Also, it is to be understood that the phraseology and terminology used herein is for the purpose of description and should not be regarded as limiting.

SUMMARY OF THE INVENTION

In one aspect, the invention provides a system and methodology for deriving, preparing and administering platelet rich plasma and adult mesenchymal autologous stem cells suitable for regenerating tissue in a living organism. The system comprises a primary container capable of removing blood from a living organism, a sealed secondary container containing a separation medium mixed with the receiving blood. The system further comprises a collecting device with a housing and a member wherein the member is moveable in a distal direction is useful in collecting and/or dispensing medium. The system further has a separation medium capable of separating red blood cells from plasma when the container contains blood and is centrifuged. The system also has a sealed or unsealed container with a transfer device designed to remove the platelet rich plasma and the adult mesenchymal autologous stem cells from the blood. The transfer device has an apparatus adapted to deliver and/or puncture a fourth container in order to provide fluid communication between the third and fourth containers. The system also has a transfer device having an apparatus to deliver the platelet rich plasma and adult mesenchymal autologous stem cells to the living organism.

In another aspect, the invention provides another system for deriving, preparing and administering platelet rich plasma and adult mesenchymal autologous stem cells suitable for regenerating tissue in a living organism. The system comprises a primary container capable of removing blood from a living organism. The system further comprises a secondary container

capable of receiving blood from the primary container and having a single opening configured to receive a cap to form a seal therein. The system further comprises a cap capable of being removed from the container and/or pierced in sealing the container for delivery of blood into the secondary container. The system also comprises a secondary container containing a separation medium capable of separating red blood cells from plasma when the container is centrifuged. The system further comprises a collecting device with a housing and a member wherein the member is moveable in a distal direction is useful in collecting and/or dispensing medium, a separation medium capable of separating red blood cells from platelet rich plasma and adult mesenchymal autologous stem cells when the container contains blood and is centrifuged. The system also includes a sealed or unsealed container with a transfer device designed to remove the platelet rich plasma and the adult mesenchymal autologous stem cells from the blood. The system also comprises a device configured to pierce the cap or in lieu of removing the cap, transfer a portion of the platelet rich plasma and adult mesenchymal autologous stem cells to a device. The system also includes a device configured to pierce the cap and transfer a portion of the platelet rich plasma and the adult mesenchymal autologous stem cells from the secondary container to the third container via pressure differentiation upon establishment of fluid communication between the cannula of the third container undergoing removal of the platelet rich plasma and the adult mesenchymal autologous stem cells until only the portion of the red blood cells remains in the secondary container.

In another aspect, the invention provides a method for deriving, preparing and administering platelet rich plasma and adult mesenchymal autologous stem cells suitable for regenerating tissue in a living organism. The method comprises drawing blood from a patient through a primary container into a secondary container containing an anti-coagulant. The blood is transferred from a secondary container. The blood is centrifuged allowing a separation of the platelet rich plasma and the adult mesencymal autologous stem cells from the whole blood. A collecting device with a housing and a member wherein the member is moveable in a distal direction is utilized in collecting the medium. The collecting device is then able to dispense its medium into an area that includes but is not limited to; musculoskeletal, ligament, tendon, cartilage, meniscus, and disc of a living organism for regeneration of said area.

In another aspect, the invention provides a method for deriving, preparing and administering platelet rich plasma and adult mesenchymal autologous stem cells suitable for regenerating tissue in a living organism. The method comprises drawing blood from a patient through a primary container into a secondary container containing an anti-coagulant. The blood is transferred from a secondary container. The blood is centrifuged allowing a separation of the platelet rich plasma and the adult mesencymal autologous stem cells from the whole blood. A collecting device with a housing and a member wherein the member is moveable in a distal direction is utilized in collecting the medium. The collecting device with a housing and a member wherein the member is moveable in a distal direction, with its contents, is able to dispense its medium into a second

medium housed in a container for multiplying, regenerating, duplicating and incubating adult mesencymal autologous stem cells for future use.

In another aspect, the invention provides a method for deriving, preparing and administering platelet rich plasma and adult mesenchymal autologous stem cells suitable for regenerating tissue in a living organism. The method comprises drawing blood from a patient through a primary container into a secondary container containing an anti-coagulant. The blood is transferred from a secondary container. The blood is centrifuged allowing a separation of the platelet rich plasma and the adult mesencymal autologous stem cells from the whole blood. A collecting device with a housing and a member wherein the member is moveable in a distal direction is utilized in collecting the medium. The collecting device with a housing and a member wherein the member is moveable in a distal direction, with its contents, is able to dispense its medium into a separate container that contains but is not limited to sodium chloride or similar solution. The container containing the medium of platelet rich plasma, adult mesenchymal autologous stem cells and sodium chloride or similar medium can now be dispensed into a living organism via intravenous or intra-arterial.

In another aspect, the invention provides a method for deriving, preparing and administering platelet rich plasma and adult mesenchymal autologous stem cells suitable for regenerating tissue in a living organism. The method comprises drawing blood from a patient through a primary container into a secondary container containing an anti-coagulant. The blood is transferred from a secondary container. The blood is centrifuged allowing a separation of the platelet rich plasma and the adult mesencymal autologous stem cells from the whole blood. A collecting device with a housing and a member wherein the member is moveable in a distal direction is utilized in collecting the medium. The collecting device with a housing and a member wherein the member is moveable in a distal direction, with its contents, is able to dispense its medium into a separate container that contains but is not limited to sodium chloride, albuterol, or a similar solution. The container containing the medium of platelet rich plasma, adult mesenchymal autologous stem cells and sodium chloride or similar medium can now be dispensed into a living organism via inhalation.

DETAILED DESCRIPTION OF THE INVENTION

The object of the present invention is therefore to provide a system and methodology for deriving, preparing and administering platelet rich plasma and adult mesenchymal autologous stem cells suitable for regenerating tissue in a living organism without resulting in viral infections and/or rejection cases when used in surgery.

Such an object is achieved by using platelet rich plasma and adult mesenchymal autologous stem cells obtained from a patient's own blood, is more reliable with respect to the rejection and/or infection risks, with little if any side effects.

Furthermore, rejection cases are known from the literature resulting from reimplanting donored stem cells produced by using human or animal proteins in patients. Such cases are indeed due to the donored origin, with respect to the receiver organism of the donored protein being reimplanted or some of the components used for preparing it.

The system and methodology for deriving, preparing and administering platelet rich plasma and adult mesenchymal autologous stem cells according to the present invention comprises a sealed container containing an anti-coagulant. The anti-coagulant separates the platelet rich plasma and adult mesenchymal autologous stem cells from the whole blood when it is introduced into the sealed container.

The system and methodology for deriving, preparing and administering platelet rich plasma and adult mesenchymal autologous stem cells according to the present invention have the great advantage of allowing the preparation of platelet rich plasma and adult mesenchymal autologous stem cells which may be used with no risk of viral infections or rejection cases.

Another advantage of the methodology for deriving, preparing and administering platelet rich plasma and adult mesenchymal autologous stem cells according to the present invention is that it allows the preparation of platelet rich plasma and adult mesenchymal autologous stem cells from patient's plasma in a very short time as well as in the formation of liquid, membranous coatings, injectables, intravenous, intra-arterial or spray. Still another advantage of the methodology for deriving, preparing and administering platelet rich plasma and adult mesenchymal autologous stem cells according to the present invention is that it allows the preparation of platelet rich plasma and adult mesenchymal autologous stem cells to be obtained at costs proportionally lower with respect to the known systems.

Further advantages of the methodology for deriving, preparing and administering platelet rich plasma and adult mesenchymal autologous stem cells to the present invention will be evident to those skilled in the art from the following detailed description of some embodiments thereof.

Containers suitable for deriving, preparing and administering platelet rich plasma and adult mesenchymal autologous stem cells according to the present invention include a glass container for blood collection as hereinafter described in Example 1. Also glass or plastic test-tubes may be used. The preferred volume of the container is from 5 to 15 ml. with preference being 10 ml. The test-tubes have preferably a diameter ranging from 12 to 16 mm and a height ranging from 75 to 100 mm. with preference being 16mm in diameter and 100mm in height. The container should be suitably thick in order to withstand the stresses resulting from the pressure difference between its inner space and the atmosphere when it is evacuated. Hemispherical or conical bottom tubes are preferably 0.7 mm thick, flat bottom tubes 1 mm thick. The plastic containers are preferably made of transparent polyester resin, 0.2-0.8 mm thick, in order to ensure the vacuum keeping for at least 12 months after production. After the preparation, the plastic test-

tubes, are preferably introduced into a tin-foil vacuum air-tight container having a heat-sealed inner polyethylene layer in order to ensure a perfect air-tightness until the date of use.

It should be noted that the evacuation of containers or test-tubes is advisable, however not necessary for putting the present invention into practice.

The containers or test-tubes are sealed by rubber or silicon pierceable caps, being suitable to ensure the container to be perfectly air-tight and to allow the vacuum plugging before the introduction of the blood and during evacuation of the platelet rich plasma and the adult mesenchymal stem cells step.

EXAMPLES

Example 1

30 ml of venous blood were drawn from a patient according to the provisions of the qualitative standards for clinical analysis, e.g. by using VACUTAINER.RTM. sterile test-tubes by Becton-Dickinson, added with a 0.106 M sodium citrate solution. For this purpose also test-tubes added with disodium or dipotassium ethylenediaminetetraacetate can be used. The sample was carefully kept sterile during the blood drawing. Finally, the sample was gently shaken for wholly mixing the components, thereby ensuring the anticoagulating action of sodium citrate. The test-tube was then introduced in a suitable centrifuge, while carefully balancing the rotor weight in order to prevent the same centrifuge to be damaged. Once the lid is sealed, the sample was centrifuged at 3500 rpm for 15 minutes, thereby separating the red cells (being thicker) from the citrated plasma (supernatant). In this case the plasma yield, mainly depending upon the characteristics of the donor blood, was as high as 65%. The test-tube containing the separated plasma was kept plugged in sterile conditions and was placed vertically in a stand for recovering the plasma itself, in this step care was taken not to shake the test-tube, in order to prevent the mixing of the two phases separated in the centrifugation. The outer portion of the test-tube cap was then sterilized by using denatured alcohol, then carefully removed. A sterile needle, being connected to a sterile syringe, was introduced in the test-tube cap. The needle was brought up to 3-4 mm apart from the separating meniscus of the two phases, and 12 ml of platelet rich plasma were drawn along with the buffy coat containing the adult mesenchymal autologous stem cells. The platelet rich plasma yield was then inserted into (4) second sterile syringes each containing 3 ml of platelet rich plasma and adult mesenchymal autologous stem cells and a sterile needle of choice was connected to each sterile syringe containing the platelet rich plasma and adult mesenchymal autologous stem cells, now ready to be immediately used.

Example 2

Approximately 30 ml of venous blood was drawn from a normotype 19 years-old patient presenting a tear of the left medial meniscus by using 10 ml sodium citrate VACUTAINER.RTM. test-tubes by Kendall, taking care to shake gently just after the drawing of the sample. The so taken blood was immediately subjected to centrifugation (15 min. at 2500 rpm) to separate the platelet rich plasma and adult mesenchymal autologous stem cells. The platelet rich plasma and adult mesenchymal autologous stem cells (12 ml) with all sterility precautions were carefully transferred into four 3 ml sterile syringes. Four, 27 gauge 1 ¼ sterile needles were attached to the sterile syringes containing the platelet rich plasma and adult mesenchymal autologous stem cells. After anesthetizing the area and after all sterility precautions, (12 ml) of platelet rich plasma and adult mesenchymal autologous stem cells were injected into the medial and lateral mensiscus of the patient's left knee. The procedure was performed approximately every three (3) weeks for 7 months and then every (4) weeks for an additional 5 months. MRI 12 months after proved the healing and regeneration of the medial and lateral meniscus, with a better post-operative course than with traditional methods.

Example 3

To produce 6 ml of platelet rich plasma and adult mesenchymal autologous stem cells, obtained as in Example 2, were transferred, with all the measures in order to preserve sterility, into two 3ml syringes according to the present invention, prepared as described in Example 1, from venous blood drawn from a normotype 30 year-old patient with chronic sprain of right ankle involving the anterior and posterior tibiofibular ligaments; the talonavicular ligament and the posterior talofibular ligament. The platelet rich plasma and adult mesenchymal autologous stem cells (6 ml) with all sterility precautions were carefully transferred into four 3 ml sterile syringes. Four, 27 gauge 1 ½ sterile needles were attached to the sterile syringes containing the platelet rich plasma and adult mesenchymal autologous stem cells. After anesthetizing the area and after all sterility precautions, (6 ml) of platelet rich plasma and adult mesenchymal autologous stem cells were injected into the right ankle as follows: 1.5ml injected into the attachment site of the distal posterior talofibular ligament; 1.5ml injected into the attachment site of the distal medial malleolus; 1.5ml injected into the distal anterior talofibular ligament; 1.5ml injected into the posterior talar process; 1.5ml injected into the sinus tarsi; and 1.5ml injected into the long axis of the metatarsal. The procedure was performed approximately every three (3) weeks for 7 months. MRI 7 months after proved the healing and regeneration of the distal posterior talofibular ligament, distal anterior talofibular ligament and the talocancaneal ligament, with a better post-operative course than with traditional methods.

Example 4

To produce 9 ml of platelet rich plasma and adult mesenchymal autologous stem cells, obtained as in Example 2, were transferred, with all the measures in order to preserve sterility, into three 3ml syringes according to the present invention, prepared as described in Example 1, from venous blood drawn from a normotype 67 year-old patient presenting a right acetabular labrum tear. The platelet rich plasma and adult mesenchymal autologous stem cells (9 ml) with all sterility precautions were carefully transferred into three 3 ml sterile syringes. Three, 25 gauge 2" sterile needles were attached to the sterile syringes containing the platelet rich plasma and adult mesenchymal autologous stem cells. After anesthetizing the area and after all sterility precautions, (9 ml) of platelet rich plasma and adult mesenchymal autologous stem cells were injected into the right hip as follows: 3ml injected into the attachment site of the iliofemoral ligament into the synovial membrane above the greater trochanter; 3ml injected into the attachment site of the iliofemoral ligament into the synovial membrane anterior to the greater trochanter; 3ml injected into the attachment site of the iliofemoral ligament into the synovial membrane posterior to the greater trochanter. The procedure was performed approximately every three (3) weeks for 7 months and then every (4) weeks for an additional 5 months. MRI 12 months after proved marked healing and regeneration of the acetabular labrum and articular cartilage as well as the transverse ligament of the acetabulum, with a better post-operative course than with traditional methods.

Example 5

To produce 9 ml of platelet rich plasma and adult mesenchymal autologous stem cells, obtained as in Example 2, were transferred, with all the measures in order to preserve sterility, into three 3ml syringes according to the present invention, prepared as described in Example 1, from venous blood drawn from a normotype 58 year-old patient presenting a SLAP TEAR (superior labrial tear anterior to posterior) of the left shoulder. The platelet rich plasma and adult mesenchymal autologous stem cells (9 ml) with all sterility precautions were carefully transferred into three, 3 ml sterile syringes. Three, 25 gauge 2" sterile needles were attached to the sterile syringes containing the platelet rich plasma and adult mesenchymal autologous stem cells. After anesthetizing the area and after all sterility precautions, (9 ml) of platelet rich plasma and adult mesenchymal autologous stem cells were injected into the left shoulder as follows: 3ml injected into the site of the anterior glenoidal labrum; 3ml injected into the site of posterior glenoidal labrum; 3ml injected into the site of the superior aspect of the labrum. The procedure was performed approximately every three (3) weeks for 7 months and then every (4) weeks for an additional 5 months. MRI 12 months after proved marked healing and regeneration of the suerior labrum and labrum anteriorly and posteriorly, with a better post-operative course than with traditional methods.

Example 6

To produce 6 ml of platelet rich plasma and adult mesenchymal autologous stem cells, obtained as in Example 2, were transferred, with all the measures in order to preserve sterility, into two 3ml syringes according to the present invention, prepared as described in Example 1, from venous blood drawn from a normotype 34 year-old patient presenting a partial tear of the right lateral collateral ligament. The platelet rich plasma and adult mesenchymal autologous stem cells (6 ml) with all sterility precautions were carefully transferred into two, 3 ml sterile syringes. Two, 27 gauge 11/4" sterile needles were attached to the sterile syringes containing the platelet rich plasma and adult mesenchymal autologous stem cells. After anesthetizing the area and after all sterility precautions, (6 ml) of platelet rich plasma and adult mesenchymal autologous stem cells were injected into the right lateral collateral and right medial collateral ligaments as follows: 1.5ml injected into the site of the ulnar collateral ligament lateral component; 1.5 ml injected into the site of the ulnar collateral ligament posterior component; 1.51 injected into the site of the radial collateral ligament; 1.5ml injected into the site of the annular ligament. The procedure was performed approximately every three (3) weeks for 7 months and then every (4) weeks for an additional 5 months. MRI 12 months after proved marked healing and regeneration of the right lateral collateral ligament, with a better post-operative course than with traditional methods.

Example 7

To produce 6 ml of platelet rich plasma and adult mesenchymal autologous stem cells, obtained as in Example 2, were transferred, with all the measures in order to preserve sterility, into two 3ml syringes according to the present invention, prepared as described in Example 1, from venous blood drawn from a normotype 45 year-old patient presenting degeneration of the cervical vertebra disc and associated cervical facets with particular interest to C4-C5-C6 bilaterally. The platelet rich plasma and adult mesenchymal autologous stem cells (6 ml) with all sterility precautions were carefully transferred into two, 3 ml sterile syringes. Two, 27 gauge 1" sterile needles were attached to the sterile syringes containing the platelet rich plasma and adult mesenchymal autologous stem cells. After anesthetizing the area and after all sterility precautions, (6 ml) of platelet rich plasma and adult mesenchymal autologous stem cells were injected into the cervical facets of C4-C5-C6 as follows: 1ml injected into the site of C4 facet on the right, followed by 1 ml injected into the site of C4 facet on the left, followed by 1ml injected into the site of C5 facet on the right, followed by 1ml injected into the site of C5 facet on the left, followed by 1ml injected into the site of C6 facet on the right, followed by 1ml injected into the C6 facet on the left. The procedure was performed approximately every three (3) weeks for 7 months and then every (4) weeks for an additional 5 months. MRI 12 months after proved marked healing and regeneration of the C4-C5-C6 facets bilaterally, with a better post-operative course than with traditional methods.

Further advantages for deriving, preparing and administering platelet rich plasma and adult mesenchymal autologous stem cells according to the present invention will be evident to those skilled in the art from the following detailed description of some embodiments thereof and may be utilized on any and all facets of the cervical region.

Example 8

To produce 6 ml of platelet rich plasma and adult mesenchymal autologous stem cells, obtained as in Example 2, were transferred, with all the measures in order to preserve sterility, into two 3ml syringes according to the present invention, prepared as described in Example 1, from venous blood drawn from a normotype 47 year-old patient presenting degeneration of the thoracic vertebra disc and associated thoracic facets with particular interest to T10-T11-12 bilaterally. The platelet rich plasma and adult mesenchymal autologous stem cells (6 ml) with all sterility precautions were carefully transferred into two, 3 ml sterile syringes. Two, 27 gauge 11/4" sterile needles were attached to the sterile syringes containing the platelet rich plasma and adult mesenchymal autologous stem cells. After anesthetizing the area and after all sterility precautions, (6 ml) of platelet rich plasma and adult mesenchymal autologous stem cells were injected into the thoracic facets of T10-T11-T12 as follows: 1ml injected into the site of T10 facet on the right, followed by 1 ml injected into the site of T10 facet on the left, followed by 1ml injected into the site of T11 facet on the right, followed by 1ml injected into the site of T11 facet on the left, followed by 1ml injected into the site of T12 facet on the right, followed by 1ml injected into the T12 facet on the left. The procedure was performed approximately every three (3) weeks for 7 months and then every (4) weeks for an additional 5 months. MRI 12 months after proved marked healing and regeneration of the T10-T11-T12 facets bilaterally, with a better post-operative course than with traditional methods.

Further advantages for deriving, preparing and administering platelet rich plasma and adult mesenchymal autologous stem cells according to the present invention will be evident to those skilled in the art from the following detailed description of some embodiments thereof and may be utilized on any and all facets of the thoracic region.

Example 9

To produce 12 ml of platelet rich plasma and adult mesenchymal autologous stem cells, obtained as in Example 2, were transferred, with all the measures in order to preserve sterility, into four 3ml syringes according to the present invention, prepared as described in Example 1, from venous blood drawn from a normotype 67 year-old patient presenting degeneration of the lumbosacral vertebra disc and associated lumbar facets with particular interest to L4-L5-S1 bilaterally. The platelet rich plasma and adult mesenchymal autologous stem cells (12 ml) with all sterility precautions were carefully transferred into four, 3 ml sterile syringes. Four, 27 gauge 2" sterile

needles were attached to the sterile syringes containing the plasma and adult mesenchymal autologous stem cells. After anesthetizing the area and after all sterility precautions, (12 ml) of platelet rich plasma and adult mesenchymal autologous stem cells were injected into the lumbosacral facets of L4-L5-S1 as follows: 2ml injected into the site of L4 facet on the right, followed by 2 ml injected into the site of L5 facet on the right, followed by 2ml injected into the site of L5 facet on the left, followed by 2ml injected into the S1 facet on the left. The procedure was performed approximately every three (3) weeks for 7 months and then every (4) weeks for an additional 5 months. MRI 12 months after proved marked healing and regeneration of the L4-L5-S1 facets bilaterally, with a better post-operative course than with traditional methods.

Further advantages for deriving, preparing and administering platelet rich plasma and adult mesenchymal autologous stem cells according to the present invention will be evident to those skilled in the art from the following detailed description of some embodiments thereof and may be utilized on any and all facets of the lumbar region.

Example 10

To produce 52 ml of platelet rich plasma and adult mesenchymal autologous stem cells, obtained as in Example 2, were transferred, with all the measures in order to preserve sterility, into (14) fourteen 3ml syringes according to the present invention, prepared as described in Example 1, from venous blood drawn from a normotype 59 year-old patient presenting COPD and sarcodosis of related to the lungs and pulmonary system. The platelet rich plasma and adult mesenchymal autologous stem cells (52 ml) with all sterility precautions were carefully transferred into fourteen, 3 ml sterile syringes. Fourteen, 21 gauge 1" sterile needles were attached to the sterile syringes containing the platelet rich plasma and adult mesenchymanl autologous stem cells. Using a nebulizer, 3ml of albuterol, its equivalent, or 3ml of .9% NaCL is placed into the medication. (1) one syringe containing 3ml of platelet rich plasma and adult mesenchymal autologous stem cells is then injected through the 21 gauge 1" needle into the medication cup. The inhalation top is then placed on the medication cup and turned clockwise until securely closed. The container is then gently shaken, to complete dissolution. The desired inhalation accessory (mouthpiece or nosepiece) is then attached to the nebulizer, the power is turned on, the nebulizer begins and the aerosol containing the adult mesenchymal autologous stem cells is generated. Inhalation treatment of platelet rich plasma and adult mesenchymal autologous stem cells was performed for approximately 10-20 minutes each session. The procedure was performed daily for (14) fourteen days.

At the end of (2) two weeks, to produce 42 ml of platelet rich plasma and adult mesenchymal autologous stem cells, the above procedure was repeated. The procedure was performed every other day for (28) days.

At the end of (4) four weeks, to produce 84 ml of platelet rich plasma and adult mesenchymal autologous stem cells, the above procedure was repeated. The procedure was performed every third day for (56) days; using a nebulizer, 3ml of albuterol, its equivalent, or 3ml of .9% NaCL was placed into the medication. (1) one syringe containing 3ml of platelet rich plasma and adult mesenchymal autologous stem cells was then injected through the 21 gauge 1" needle into the medication cup. The inhalation top was then placed on the medication cup and turned clockwise until securely closed. The container was then gently shaken, to complete dissolution. The desired inhalation accessory (mouthpiece or nosepiece) was then attached to the nebulizer, the power was turned on, the nebulizer began and the aerosol containing the adult mesenchymal autologous stem cells was generated. Inhalation treatment of adult mesenchymal autologous stem cells was performed for approximately 10-20 minutes each session. The procedure was performed every third day for (56) fifty-six days.

At the end of (8) eight weeks, to produce 96 ml of platelet rich plasma and adult mesenchymal autologous stem cells, the above procedure was repeated, placing 3ml of the platelet rich plasma and adult mesenchymal autologous stem cells into (21) twenty-one separate 3ml sterile syringes, each with (1) 1" sterile needle attached to syringe. The procedure was performed every fourth day for (84) days using a nebulizer, 3ml of albuterol, its equivalent, or 3ml of .9% NaCL was placed into the medication. (1) one syringe containing 3ml of platelet rich plasma and adult mesenchymal autologous stem cells was then injected through the 21 gauge 1" needle into the medication cup. The inhalation top was then placed on the medication cup and turned clockwise until securely closed. The container was then gently shaken, to complete dissolution. The desired inhalation accessory (mouthpiece or nosepiece) was then attached to the nebulizer, the power was turned on, the nebulizer began and the aerosol containing the platelet rich plasma and adult mesenchymal autologous stem cells was generated. Inhalation treatment of platelet rich plasma and adult mesenchymal autologous stem cells was performed for approximately 10-20 minutes each session. The procedure was performed every fourth day for (84) eighty-four days.

At the end of (12) twelve weeks, to produce 51 ml of platelet rich plasma and adult mesenchymal autologous stem cells, the above procedure was repeated, placing 3ml of the platelet rich plasma and adult mesenchymal autologous stem cells into (17)seventeen separate 3ml sterile syringes, each with (1) 1" sterile needle attached to syringe. The procedure was performed every fifth day for (84) days using a nebulizer, 3ml of albuterol, its equivalent, or 3ml of .9% NaCL was placed into the medication. (1) one syringe containing 3ml of platelet rich plasma and adult mesenchymal autologous stem cells was then injected through the 21 gauge 1" needle into the medication cup. The inhalation top was then placed on the medication cup and turned clockwise until securely closed. The container was then gently shaken, to complete dissolution. The desired inhalation accessory (mouthpiece or nosepiece) was then attached to the nebulizer, the power was turned on, the nebulizer began and the aerosol containing the platelet rich plasma and adult mesenchymal autologous stem cells was generated. Inhalation treatment of platelet rich

plasma and adult mesenchymal autologous stem cells was performed for approximately 10-20 minutes each session. The procedure was performed every fifth day for (84) eighty-four days.

At the end of (12) twelve weeks, to produce 42 ml of platelet rich plasma and adult mesenchymal autologous stem cells, the above procedure was repeated, placing 3ml of the platelet rich plasma and adult mesenchymal autologous stem cells into (14) fourteen separate 3ml sterile syringes, each with (1) 1" sterile needle attached to syringe. The procedure was performed every sixth day for (84) days using a nebulizer, 3ml of albuterol, its equivalent, or 3ml of .9% NaCL was placed into the medication. (1) one syringe containing 3ml of platelet rich plasma and adult mesenchymal autologous stem cells was then injected through the 21 gauge 1" needle into the medication cup. The inhalation top was then placed on the medication cup and turned clockwise until securely closed. The container was then gently shaken, to complete dissolution. The desired inhalation accessory (mouthpiece or nosepiece) was then attached to the nebulizer, the power was turned on, the nebulizer began and the aerosol containing the platelet rich plasma and adult mesenchymal autologous stem cells was generated. Inhalation treatment of platelet rich plasma and adult mesenchymal autologous stem cells was performed for approximately 10-20 minutes each session. The procedure was performed every sixth day for (84) eighty-four days. 12 months after proved marked healing and regeneration of the lungs and pulmonary system, with a better post-operative course than with traditional methods.

Further advantages for deriving, preparing and administering platelet rich plasma and adult mesenchymal autologous stem cells according to the present invention will be evident to those skilled in the art from the following detailed description of some embodiments thereof and may be utilized on any and all systems of a living organism.

Example 11

To produce 42 ml of platelet rich plasma and adult mesenchymal autologous stem cells, obtained as in Example 2, were transferred, with all the measures in order to preserve sterility, into (14) fourteen 3ml syringes according to the present invention, prepared as described in Example 1, from venous blood drawn from a normotype 70 year-old patient presenting aortic stenosis and mitral valve regurgitation related to the heart and cardiovascular system. The platelet rich plasma and adult mesenchymal autologous stem cells (42 ml) with all sterility precautions were carefully transferred into fourteen, 3 ml sterile syringes. Fourteen, 21 gauge 1" sterile needles were attached to the sterile syringes containing the platelet rich plasma and adult mesenchymal autologous stem cells. Using a nebulizer, 3ml of albuterol, its equivalent, or 3ml of .9% NaCL is placed into the medication. (1) one syringe containing 3ml of platelet rich plasma and adult mesenchymal autologous stem cells is then injected through the 21 gauge 1" needle into the medication cup. The inhalation top is then placed on the medication cup and turned clockwise until securely closed. The container is then gently shaken, to complete dissolution. The desired

inhalation accessory (mouthpiece or nosepiece) is then attached to the nebulizer, the power is turned on, the nebulizer begins and the aerosol containing the adult mesenchymal autologous stem cells is generated. Inhalation treatment of platelet rich plasma and adult mesenchymal autologous stem cells was performed for approximately 10-20 minutes each session. The procedure was performed daily for (14) fourteen days.

At the end of (2) two weeks, to produce 42 ml of platelet rich plasma and adult mesenchymal autologous stem cells, the above procedure was repeated. The procedure was performed every other day for (28) days.

At the end of (4) four weeks, to produce 84 ml of platelet rich plasma and adult mesenchymal autologous, the above procedure was repeated. The procedure was performed every third day for (56) days; using a nebulizer, 3ml of albuterol, its equivalent, or 3ml of .9% NaCL was placed into the medication. (1) one syringe containing 3ml of platelet rich plasma and adult mesenchymal autologous stem cells was then injected through the 21 gauge 1" needle into the medication cup. The inhalation top was then placed on the medication cup and turned clockwise until securely closed. The container was then gently shaken, to complete dissolution. The desired inhalation accessory (mouthpiece or nosepiece) was then attached to the nebulizer, the power was turned on, the nebulizer began and the aerosol containing the platelet rich plasma and adult mesenchymal autologous stem cells was generated. Inhalation treatment of platelet rich plasma and adult mesenchymal autologous stem cells was performed for approximately 10-20 minutes each session. The procedure was performed every third day for (56) fifty-six days.

At the end of (8) eight weeks, to produce 96 ml of platelet rich plasma and adult mesenchymal autologous stem cells, the above procedure was repeated, placing 3ml of the platelet rich plasma and adult mesenchymal autologous stem cells into (21) twenty-one separate 3ml sterile syringes, each with (1) 1" sterile needle attached to syringe. The procedure was performed every fourth day for (84) days using a nebulizer, 3ml of albuterol, its equivalent, or 3ml of .9% NaCL was placed into the medication. (1) one syringe containing 3ml of platelet rich plasma and adult mesenchymal autologous stem cells was then injected through the 21 gauge 1" needle into the medication cup. The inhalation top was then placed on the medication cup and turned clockwise until securely closed. The container was then gently shaken, to complete dissolution. The desired inhalation accessory (mouthpiece or nosepiece) was then attached to the nebulizer, the power was turned on, the nebulizer began and the aerosol containing the adult mesenchymal autologous stem cells was generated. Inhalation treatment of platelet rich plasma and adult mesenchymal autologous stem cells was performed for approximately 10-20 minutes each session. The procedure was performed every fourth day for (84) eighty-four days.

At the end of (12) twelve weeks, to produce 51 ml of platelet rich plasma and adult mesenchymal autologous stem cells, the above procedure was repeated, placing 3ml of the platelet rich plasma and adult mesenchymal autologous stem cells into (17)seventeen separate 3ml sterile syringes, each with (1) 1" sterile needle attached to syringe. The procedure was performed every fifth day

for (84) days using a nebulizer, 3ml of albuterol, its equivalent, or 3ml of .9% NaCL was placed into the medication. (1) one syringe containing 3ml of platelet rich plasma and adult mesenchymal autologous stem cells was then injected through the 21 gauge 1" needle into the medication cup. The inhalation top was then placed on the medication cup and turned clockwise until securely closed. The container was then gently shaken, to complete dissolution. The desired inhalation accessory (mouthpiece or nosepiece) was then attached to the nebulizer, the power was turned on, the nebulizer began and the aerosol containing the adult mesenchymal autologous stem cells was generated. Inhalation treatment of adult mesenchymal autologous stem cells was performed for approximately 10-20 minutes each session. The procedure was performed every fifth day for (84) eighty-four days.

At the end of (12) twelve weeks, to produce 42 ml of platelet rich plasma and adult mesenchymal autologous stem cells, the above procedure was repeated, placing 3ml of the platelet rich plasma and adult mesenchymal autologous stem cells into (14) fourteen separate 3ml sterile syringes, each with (1) 1" sterile needle attached to syringe. The procedure was performed every sixth day for (84) days using a nebulizer, 3ml of albuterol, its equivalent, or 3ml of .9% NaCL was placed into the medication. (1) one syringe containing 3ml of platelet rich plasma and adult mesenchymal autologous stem cells was then injected through the 21 gauge 1" needle into the medication cup. The inhalation top was then placed on the medication cup and turned clockwise until securely closed. The container was then gently shaken, to complete dissolution. The desired inhalation accessory (mouthpiece or nosepiece) was then attached to the nebulizer, the power was turned on, the nebulizer began and the aerosol containing the adult mesenchymal autologous stem cells was generated. Inhalation treatment of platelet rich plasma and adult mesenchymal autologous stem cells was performed for approximately 10-20 minutes each session. The procedure was performed every sixth day for (84) eighty-four days. 12 months after proved marked healing and regeneration of the mitral and aortic valve with no gurgling or stenosis on examination and with a better post-operative course than with traditional methods.

Further advantages for deriving, preparing and administering platelet rich plasma and adult mesenchymal autologous stem cells according to the present invention will be evident to those skilled in the art from the following detailed description of some embodiments thereof and may be utilized on any and all systems of a living organism.

Example 12

To produce 48 ml of platelet rich plasma and adult mesenchymal autologous stem cells, obtained as in Example 2, were transferred, with all the measures in order to preserve sterility, into (16) sixteen 3ml syringes according to the present invention, prepared as described in Example 1, from venous blood drawn from a normotype 52 year-old patient presenting T.I.A. as a result of Lyme Disease and autoimmune scleroderma. The platelet rich plasma and adult mesenchymal

autologous stem cells (48 ml) with all sterility precautions were carefully transferred into sixteen, 3 ml sterile syringes. Sixteen, 21 gauge 1" sterile needles were attached to the sterile syringes containing the platelet rich plasma and adult mesenchymal autologous stem cells. Using a nebulizer, 3ml of albuterol, its equivalent, or 3ml of .9% NaCL is placed into the medication. (1) one syringe containing 3ml of platelet rich plasma and adult mesenchymal autologous stem cells is then injected through the 21 gauge 1" needle into the medication cup. The inhalation top is then placed on the medication cup and turned clockwise until securely closed. The container is then gently shaken, to complete dissolution. The desired inhalation accessory (mouthpiece or nosepiece) is then attached to the nebulizer, the power is turned on, the nebulizer begins and the aerosol containing the platelet rich plasma and adult mesenchymal autologous stem cells is generated. Inhalation treatment of platelet rich plasma and adult mesenchymal autologous stem cells was performed for approximately 10-20 minutes each session. The procedure was performed daily for (14) fourteen days with the exception of days (7) seven and (14) fourteen.

Day (7) seven, and day (14) fourteen, (6) six ml of platelet rich plasma and adult mesenchymal autologous stem cells were injected into an IV bag containing 250ml of .9% NaCL at room temperature and gently shaken to complete dissolution. The solution was then injected intravenously into the patient for approximately (45) forty-five minutes.

At the end of (2) two weeks, to produce 51 ml of platelet rich plasma and adult mesenchymal autologous stem cells, the above procedure was repeated. The procedure was performed every other day for (28) days with the exception of days (21) twenty-one, (28) twenty-eight and (42) forty-two, (6) six ml of platelet rich plasma and adult mesenchymal autologous stem cells were injected into an IV bag containing 250ml of .9% NaCL at room temperature and gently shaken to complete dissolution. The solution was then injected intravenously into the patient for approximately (45) forty-five minutes at each session.

At the end of (4) four weeks, to produce 96 ml of platelet rich plasma and adult mesenchymal autologous stem cells, the above procedure was repeated, placing 3ml of the platelet rich plasma and adult mesenchymal autologous stem cells into (32) thirty-two separate 3ml sterile syringes, each with (1) 1" sterile needle attached to syringe. The procedure was performed every third day for (84) days with the exception of days (63) sixty-three, (84) eighty-four, (105) one hundred five and (126) one hundred twenty-six; (6) six ml of platelet rich plasma and adult mesenchymal autologous stem cells were injected into an IV bag containing 250ml of .9% NaCL at room temperature and gently shaken to complete dissolution. The solution was then injected intravenously into the patient for approximately (45) forty-five minutes at each session.

At the end of (12) twelve weeks, to produce 72 ml of platelet rich plasma and adult mesenchymal autologous stem cells, the above procedure was repeated, placing 3ml of the platelet rich plasma and adult mesenchymal autologous stem cells into (24) twenty-four separate 3ml sterile syringes, each with (1) 1" sterile needle attached to syringe. The procedure was performed every fourth day for (84) days with the exception of days (154) one hundred fifty-four, (182) one hundred

eighty-two and (210) two hundred ten, (6) six ml of platelet rich plasma and adult mesenchymal autologous stem cells were injected into an IV bag containing 250ml of .9% NaCL at room temperature and gently shaken to complete dissolution. The solution was then injected intravenously into the patient for approximately (45) forty-five minutes at each session.

At the end of (12) twelve weeks, to produce 56 ml of platelet rich plasma and adult mesenchymal autologous stem cells, the above procedure was repeated, placing 3ml of the platelet rich plasma and adult mesenchymal autologous stem cells into (19) nineteen separate 3ml sterile syringes, each with (1) 1" sterile needle attached to syringe. The procedure was performed every fifth day for (84) days with the exception of days (252) two hundred fifty-two and (294) two hundred ninety-four, (6) six ml of platelet rich plasma and adult mesenchymal autologous stem cells were injected into an IV bag containing 250ml of .9% NaCL at room temperature and gently shaken to complete dissolution. The solution was then injected intravenously into the patient for approximately (45) forty-five minutes at each session.

At the end of (12) twelve weeks, to produce 48 ml of platelet rich plasma and adult mesenchymal autologous stem cells, the above procedure was repeated, placing 3ml of the platelet rich plasma and adult mesenchymal autologous stem cells into (16) sixteen separate 3ml sterile syringes, each with (1) 1" sterile needle attached to syringe. The procedure was performed every sixth day for (84) days with the exception of days (336) three hundred thirty-six and (379) three hundred seventy-nine, (6) six ml of platelet rich plasma and adult mesenchymal autologous stem cells were injected into an IV bag containing 250ml of .9% NaCL at room temperature and gently shaken to complete dissolution. The solution was then injected intravenously into the patient for approximately (45) forty-five minutes at each session.

Further advantages for deriving, preparing and administering platelet rich plasma and adult mesenchymal autologous stem cells according to the present invention will be evident to those skilled in the art from the following detailed description of some embodiments thereof and may be utilized on any and all systems of a living organism.

Example 13

To produce 48 ml of platelet rich plasma and adult mesenchymal autologous stem cells, obtained as in Example 2, were transferred, with all the measures in order to preserve sterility, into (16) sixteen 3ml syringes according to the present invention, prepared as described in Example 1, from venous blood drawn from a normotype 38 year-old patient presenting multiple sclerosis with a history of (10) ten years. The platelet rich plasma and adult mesenchymal autologous stem cells (48 ml) with all sterility precautions were carefully transferred into sixteen, 3 ml sterile syringes. Sixteen, 21 gauge 1" sterile needles were attached to the sterile syringes containing the platelet rich plasma and adult mesenchymal autologous stem cells. Using a nebulizer, 3ml of albuterol, its equivalent, or 3ml of .9% NaCL is placed into the medication. (1) one syringe

containing 3ml of plasma and adult mesenchymal autologous stem cells is then injected through the 21 gauge 1" needle into the medication cup. The inhalation top is then placed on the medication cup and turned clockwise until securely closed. The container is then gently shaken, to complete dissolution. The desired inhalation accessory (mouthpiece or nosepiece) is then attached to the nebulizer, the power is turned on, the nebulizer begins and the aerosol containing the platelet rich plasma and adult mesenchymal autologous stem cells is generated. Inhalation treatment of platelet rich plasma and adult mesenchymal autologous stem cells was performed for approximately 10-20 minutes each session. The procedure was performed daily for (14) fourteen days with the exception of days (7) seven and (14) fourteen.

Day (7) seven, and day (14) fourteen, (6) six ml of platelet rich plasma and adult mesenchymal autologous stem cells were injected into an IV bag containing 250ml of .9% NaCL at room temperature and gently shaken to complete dissolution. The solution was then injected intravenously into the patient for approximately (45) forty-five minutes.

At the end of (2) two weeks, to produce 51 ml of platelet rich plasma and adult mesenchymal autologous stem cells, the above procedure was repeated. The procedure was performed every other day for (28) days with the exception of days (21) twenty-one, (28) twenty-eight and (42) forty-two, (6) six ml of platelet rich plasma and adult mesenchymal autologous stem cells were injected into an IV bag containing 250ml of .9% NaCL at room temperature and gently shaken to complete dissolution. The solution was then injected intravenously into the patient for approximately (45) forty-five minutes at each session.

At the end of (4) four weeks, to produce 96 ml of platelet rich plasma and adult mesenchymal autologous stem cells, the above procedure was repeated, placing 3ml of the platelet rich plasma and adult mesenchymal autologous stem cells into (32) thirty-two separate 3ml sterile syringes, each with (1) 1" sterile needle attached to syringe. The procedure was performed every third day for (84) days with the exception of days (63) sixty-three, (84) eighty-four, (105) one hundred five and (126) one hundred twenty-six; (6) six ml of platelet rich plasma and adult mesenchymal autologous stem cells were injected into an IV bag containing 250ml of .9% NaCL at room temperature and gently shaken to complete dissolution. The solution was then injected intravenously into the patient for approximately (45) forty-five minutes at each session.

At the end of (12) twelve weeks, to produce 72 ml of platelet rich plasma and adult mesenchymal autologous stem cells, the above procedure was repeated, placing 3ml of the platelet rich plasma and adult mesenchymal autologous stem cells into (24) twenty-four separate 3ml sterile syringes, each with (1) 1" sterile needle attached to syringe. The procedure was performed every fourth day for (84) days with the exception of days (154) one hundred fifty-four, (182) one hundred eighty-two and (210) two hundred ten, (6) six ml of platelet rich plasma and adult mesenchymal autologous stem cells were injected into an IV bag containing 250ml of .9% NaCL at room temperature and gently shaken to complete dissolution. The solution was then injected intravenously into the patient for approximately (45) forty-five minutes at each session.

At the end of (12) twelve weeks, to produce 56 ml of platelet rich plasma and adult mesenchymal autologous stem cells, the above procedure was repeated, placing 3ml of the adult mesenchymal autologous stem cells into (19) nineteen separate 3ml sterile syringes, each with (1) 1" sterile needle attached to syringe. The procedure was performed every fifth day for (84) days with the exception of days (252) two hundred fifty-two and (294) two hundred ninety-four, (6) six ml of platelet rich plasma and adult mesenchymal autologous stem cells were injected into an IV bag containing 250ml of .9% NaCL at room temperature and gently shaken to complete dissolution. The solution was then injected intravenously into the patient for approximately (45) forty-five minutes at each session.

At the end of (12) twelve weeks, to produce 48 ml of platelet rich plasma and adult mesenchymal autologous stem cells, the above procedure was repeated, placing 3ml of the platelet rich plasma and adult mesenchymal autologous stem cells into (16) sixteen separate 3ml sterile syringes, each with (1) 1" sterile needle attached to syringe. The procedure was performed every sixth day for (84) days with the exception of days (336) three hundred thirty-six and (379) three hundred seventy-nine, (6) six ml of platelet rich plasma and adult mesenchymal autologous stem cells were injected into an IV bag containing 250ml of .9% NaCL at room temperature and gently shaken to complete dissolution. The solution was then injected intravenously into the patient for approximately (45) forty-five minutes at each session.

Further advantages for deriving, preparing and administering platelet rich plasma and adult mesenchymal autologous stem cells according to the present invention will be evident to those skilled in the art from the following detailed description of some embodiments thereof and may be utilized on any and all systems of a living organism.

Example 14

To produce 48 ml of platelet rich plasma and adult mesenchymal autologous stem cells, obtained as in Example 2, were transferred, with all the measures in order to preserve sterility, into (16) sixteen 3ml syringes according to the present invention, prepared as described in Example 1, from venous blood drawn from a normotype 45 year-old patient presenting Parkinson's Disease with a history of (10) ten years. The platelet rich plasma and adult mesenchymal autologous stem cells (48 ml) with all sterility precautions were carefully transferred into sixteen, 3 ml sterile syringes. Sixteen, 21 gauge 1" sterile needles were attached to the sterile syringes containing the platelet rich plasma and adult mesenchymal autologous stem cells. Using a nebulizer, 3ml of albuterol, its equivalent, or 3ml of .9% NaCL is placed into the medication. (1) one syringe containing 3ml of platelet rich plasma and adult mesenchymal autologous stem cells is then injected through the 21 gauge 1" needle into the medication cup. The inhalation top is then placed on the medication cup and turned clockwise until securely closed. The container is then gently shaken, to complete dissolution. The desired inhalation accessory (mouthpiece or

nosepiece) is then attached to the nebulizer, the power is turned on, the nebulizer begins and the aerosol containing the adult mesenchymal autologous stem cells is generated. Inhalation treatment of platelet rich plasma and adult mesenchymal autologous stem cells was performed for approximately 10-20 minutes each session. The procedure was performed daily for (14) fourteen days with the exception of days (7) seven and (14) fourteen.

Day (7) seven, and day (14) fourteen, (6) six ml of platelet rich plasma and adult mesenchymal autologous stem cells were injected into an IV bag containing 250ml of .9% NaCL at room temperature and gently shaken to complete dissolution. The solution was then injected intravenously into the patient for approximately (45) forty-five minutes.

At the end of (2) two weeks, to produce 51 ml of platelet rich plasma and adult mesenchymal autologous stem cells, the above procedure was repeated. The procedure was performed every other day for (28) days with the exception of days (21) twenty-one, (28) twenty-eight and (42) forty-two, (6) six ml of platelet rich plasma and adult mesenchymal autologous stem cells were injected into an IV bag containing 250ml of .9% NaCL at room temperature and gently shaken to complete dissolution. The solution was then injected intravenously into the patient for approximately (45) forty-five minutes at each session.

At the end of (4) four weeks, to produce 96 ml of platelet rich plasma and adult mesenchymal autologous stem cells, the above procedure was repeated, placing 3ml of the platelet rich plasma and adult mesenchymal autologous stem cells into (32) thirty-two separate 3ml sterile splayringes, each with (1) 1" sterile needle attached to syringe. The procedure was performed every third day for (84) days with the exception of days (63) sixty-three, (84) eighty-four, (105) one hundred five and (126) one hundred twenty-six; (6) six ml of platelet rich plasma and adult mesenchymal autologous stem cells were injected into an IV bag containing 250ml of .9% NaCL at room temperature and gently shaken to complete dissolution. The solution was then injected intravenously into the patient for approximately (45) forty-five minutes at each session.

At the end of (12) twelve weeks, to produce 72 ml of platelet rich plasma and adult mesenchymal autologous stem cells, the above procedure was repeated, placing 3ml of the platelet rich plasma and adult mesenchymal autologous stem cells into (24) twenty-four separate 3ml sterile syringes, each with (1) 1" sterile needle attached to syringe. The procedure was performed every fourth day for (84) days with the exception of days (154) one hundred fifty-four, (182) one hundred eighty-two and (210) two hundred ten, (6) six ml of platelet rich plasma and adult mesenchymal autologous stem cells were injected into an IV bag containing 250ml of .9% NaCL at room temperature and gently shaken to complete dissolution. The solution was then injected intravenously into the patient for approximately (45) forty-five minutes at each session.

At the end of (12) twelve weeks, to produce 56 ml of platelet rich plasma and adult mesenchymal autologous stem cells, the above procedure was repeated, placing 3ml of the platelet rich plasma and adult mesenchymal autologous stem cells into (19) nineteen separate 3ml sterile syringes,

each with (1) 1" sterile needle attached to syringe. The procedure was performed every fifth day for (84) days with the exception of days (252) two hundred fifty-two and (294) two hundred ninety-four, (6) six ml of platelet rich plasma and adult mesenchymal autologous stem cells were injected into an IV bag containing 250ml of .9% NaCL at room temperature and gently shaken to complete dissolution. The solution was then injected intravenously into the patient for approximately (45) forty-five minutes at each session.

At the end of (12) twelve weeks, to produce 48 ml of platelet rich plasma and adult mesenchymal autologous stem cells, the above procedure was repeated, placing 3ml of the platelet rich plasma and adult mesenchymal autologous stem cells into (16) sixteen separate 3ml sterile syringes, each with (1) 1" sterile needle attached to syringe. The procedure was performed every sixth day for (84) days with the exception of days (336) three hundred thirty-six and (379) three hundred seventy-nine, (6) six ml of platelet rich plasma and adult mesenchymal autologous stem cells were injected into an IV bag containing 250ml of .9% NaCL at room temperature and gently shaken to complete dissolution. The solution was then injected intravenously into the patient for approximately (45) forty-five minutes at each session.

Further advantages for deriving, preparing and administering platelet rich plasma and adult mesenchymal autologous stem cells according to the present invention will be evident to those skilled in the art from the following detailed description of some embodiments thereof and may be utilized on any and all systems of a living organism.

Example 15

To produce 48 ml of platelet rich plasma and adult mesenchymal autologous stem cells, obtained as in Example 2, were transferred, with all the measures in order to preserve sterility, into (16) sixteen 3ml syringes according to the present invention, prepared as described in Example 1, from venous blood drawn from a normotype 65 year-old patient presenting COPD and macular degeneration with a history of (15) fifteeen years. The platelet rich plasma and adult mesenchymal autologous stem cells (48 ml) with all sterility precautions were carefully transferred into sixteen, 3 ml sterile syringes. Sixteen, 21 gauge 1" sterile needles were attached to the sterile syringes containing the platelet rich plasma and adult mesenchymal autologous stem cells. Using a nebulizer, 3ml of albuterol, its equivalent, or 3ml of .9% NaCL is placed into the medication. (1) one syringe containing 3ml of platelet rich plasma and adult mesenchymal autologous stem cells is then injected through the 21 gauge 1" needle into the medication cup. The inhalation top is then placed on the medication cup and turned clockwise until securely closed. The container is then gently shaken, to complete dissolution. The desired inhalation accessory (mouthpiece or nosepiece) is then attached to the nebulizer, the power is turned on, the nebulizer begins and the aerosol containing the platelet rich plasma and adult mesenchymal autologous stem cells is generated. Inhalation treatment of platelet rich plasma and adult

mesenchymal autologous stem cells was performed for approximately 10-20 minutes each session. The procedure was performed daily for (14) fourteen days with the exception of days (7) seven and (14) fourteen.

Day (7) seven, and day (14) fourteen, (6) six ml of platelet rich plasma and adult mesenchymal autologous stem cells were injected into an IV bag containing 250ml of .9% NaCL at room temperature and gently shaken to complete dissolution. The solution was then injected intravenously into the patient for approximately (45) forty-five minutes.

At the end of (2) two weeks, to produce 51 ml of platelet rich plasma and adult mesenchymal autologous stem cells, the above procedure was repeated. The procedure was performed every other day for (28) days with the exception of days (21) twenty-one, (28) twenty-eight and (42) forty-two, (6) six ml of platelet rich plasma and adult mesenchymal autologous stem cells were injected into an IV bag containing 250ml of .9% NaCL at room temperature and gently shaken to complete dissolution. The solution was then injected intravenously into the patient for approximately (45) forty-five minutes at each session.

At the end of (4) four weeks, to produce 96 ml of platelet rich plasma and adult mesenchymal autologous stem cells, the above procedure was repeated, placing 3ml of the platelet rich plasma and adult mesenchymal autologous stem cells into (32) thirty-two separate 3ml sterile syringes, each with (1) 1" sterile needle attached to syringe. The procedure was performed every third day for (84) days with the exception of days (63) sixty-three, (84) eighty-four, (105) one hundred five and (126) one hundred twenty-six; (6) six ml of platelet rich plasma and adult mesenchymal autologous stem cells were injected into an IV bag containing 250ml of .9% NaCL at room temperature and gently shaken to complete dissolution. The solution was then injected intravenously into the patient for approximately (45) forty-five minutes at each session.

At the end of (12) twelve weeks, to produce 72 ml of platelet rich plasma and adult mesenchymal autologous stem cells, the above procedure was repeated, placing 3ml of the platelet rich plasma and adult mesenchymal autologous stem cells into (24) twenty-four separate 3ml sterile syringes, each with (1) 1" sterile needle attached to syringe. The procedure was performed every fourth day for (84) days with the exception of days (154) one hundred fifty-four, (182) one hundred eighty-two and (210) two hundred ten, (6) six ml of platelet rich plasma and adult mesenchymal autologous stem cells were injected into an IV bag containing 250ml of .9% NaCL at room temperature and gently shaken to complete dissolution. The solution was then injected intravenously into the patient for approximately (45) forty-five minutes at each session.

At the end of (12) twelve weeks, to produce 56 ml of platelet rich plasma and adult mesenchymal autologous stem cells, the above procedure was repeated, placing 3ml of the platelet rich plasma and adult mesenchymal autologous stem cells into (19) nineteen separate 3ml sterile syringes, each with (1) 1" sterile needle attached to syringe. The procedure was performed every fifth day for (84) days with the exception of days (252) two hundred fifty-two and (294) two hundred

ninety-four, (6) six ml of platelet rich plasma and adult mesenchymal autologous stem cells were injected into an IV bag containing 250ml of .9% NaCL at room temperature and gently shaken to complete dissolution. The solution was then injected intravenously into the patient for approximately (45) forty-five minutes at each session.

At the end of (12) twelve weeks, to produce 48 ml of platelet rich plasma and adult mesenchymal autologous stem cells, the above procedure was repeated, placing 3ml of the platelet rich plasma and adult mesenchymal autologous stem cells into (16) sixteen separate 3ml sterile syringes, each with (1) 1" sterile needle attached to syringe. The procedure was performed every sixth day for (84) days with the exception of days (336) three hundred thirty-six and (379) three hundred seventy-nine, (6) six ml of platelet rich plasma and adult mesenchymal autologous stem cells were injected into an IV bag containing 250ml of .9% NaCL at room temperature and gently shaken to complete dissolution. The solution was then injected intravenously into the patient for approximately (45) forty-five minutes at each session.

Further advantages for deriving, preparing and administering platelet rich plasma and adult mesenchymal autologous stem cells according to the present invention will be evident to those skilled in the art from the following detailed description of some embodiments thereof and may be utilized on any and all systems of a living organism.

Example 16

To produce 36 ml of platelet rich plasma and adult mesenchymal autologous stem cells, obtained as in Example 2, were transferred, with all the measures in order to preserve sterility, into twelve 3ml syringes according to the present invention, prepared as described in Example 1, from venous blood drawn from a normotype 24 year-old patient. The platelet rich plasma and adult mesenchymal autologous stem cells (36 ml) with all sterility precautions were carefully transferred into twelve, 3 ml sterile syringes. Twelve, 21 gauge 1" sterile needles were attached to the sterile syringes containing the platelet rich plasma and adult mesenchymal autologous stem cells. 3ml of platelet rich plasma and adult mesencymal autologous stem cells with all sterility precautions were carefully placed into a petrie dish containing a serum free culture medium specifically formulated for the culture of pure populated adult mesenchymal autologous stem cells. By blocking protein membrane CD47 variations of the adult mesencymal stem cell may be grown into specific cells.

Further advantages for deriving, preparing and administering adult mesenchymal autologous stem cells according to the present invention will be evident to those skilled in the art from the following detailed description of some embodiments thereof and may be utilized on any and all systems of a living organism.

Generally speaking, the invention provides integrated systems and methodologies for deriving, preparing and administering adult mesenchymal autologous stem cells suitable for regenerating tissue in a living organism without resulting in viral infections and/or rejection cases when used in oral, injectibles, intravenously, intra-arterial, minor and major surgery.

In one embodiment (shown in FIG. 1), the system comprises a primary container 10 capable of removing blood 11 from a living organism into a secondary container 12. Preferably, the secondary container 12, are tubes, and more particularly, test tubes, although any container that is capable of holding a fluid or liquid and being centrifuged is suitable for use with the invention. Preferably, the container 12, is made from glass or plastics.

The primary container 10 must be capable of drawing blood therein using standard venipuncture techniques into secondary container 12. Preferably the secondary container 12 is sealed with a seal 13 while the blood is being drawn to prevent contamination, although the container 12 may be sealed shortly thereafter. A variety of seals 13 can be used to seal the secondary container 12, e.g., a rubber stopper, cap, foam, elastomer or other composite. The seal 13 should be capable of being pierced or punctured, and therefore rubber and silicone are preferred materials from which the seal is fabricated, although any material that provides a seal and is capable of being pierced can be used.

In another embodiment (shown in FIG. 3), the secondary container 12 may contain an anticoagulant solution 14. The anticoagulant 14 may comprise sodium citrate, ethylenelendiaminetetraacetic acid disodium salt, ethylenelendiaminetetraacetic acid dipotassium salt and tripotassium and combinations thereof. Preferably, the secondary container 12 contains a sodium citrate solution. The anticoagulant 14 tends to thin blood collected in the secondary container 12 in order to place it in condition for centrifugation.

In another embodiment, the secondary container may also contain one or more of an antibiotic, an analgesic, a cancer therapeutic, a platelet-growth factor and a bone morphogenic protein. Other therapeutic agents which can be topically administered may also be included. Examples of antibiotics include, but are not limited to, ampicillin, erythromycin and tobramycin. Analgesics include, but are not limited to, aspirin and codeine. Cancer therapeutics include, but are not limited to, 5-fluor-uracile.

Turning now to the operation of the first embodiment, once blood has been drawn into the secondary container 12 using standard venipuncture techniques, the blood is anticoagulated by the anti-coagulant 14 therein. Typically, the secondary container 12 is sealed while the blood is being drawn, however, it may be sealed thereafter. Sealing the secondary container 12 prevents contamination of the contents therein.

Thereafter, the secondary container 12 and its contents 11 (i.e. blood, anti-coagulant 14) are centrifuged. Acceptable centrifugation can take place at a gravitational force in the range of 900 to 3,500 xG for 5 to 20 minutes. In a preferred embodiment, the secondary container is centrifuged at a gravitational force of about 3,000 xG for about fifteen minutes. This initial centrifugation separates the secondary container's contents or fractions into a plurality of layers as shown, e.g., in FIG.9.

The layers include (in order from the bottom of the secondary container 12 to the top of the container after centrifugation) as shown, e.g., in FIG 9: the red blood cell layer 15, the buffy layer containing adult mesenchymal autologous stem cells 16, the platelet-rich plasma layer 17, and finally a residual gas 18 volume at a pressure equal to atmospheric. The proportions of these layers may vary from application to application, and are shown here in these proportions for illustrative purposes only. Subsequent to centrifugation, the sealed secondary holder 12 is placed into the transfer device 19 is used to house the secondary container 12 in the centrifuge.

In other words, the secondary container 12 is placed in the centrifuge such that the sealed opening is in the highest vertical position as shown in FIG. 8. Placing the secondary container in such a manner allows gravity to control the order in which the layers are arranged. Below the seal 13 are the following layers in sequence from top to bottom after centrifuge as shown, e.g., in FIG 9: residual gas18, platelet rich plasma 17, buffy layer containing adult mesenchymal autologous stem cells 16, and the red blood cells 15.

Next, the secondary container 12 is placed in a vertical position with its sealed opening 13 in the top most position as best shown in FIG. 8. This positions the secondary container 12 for the centrifuging of the secondary container's contents therein. FIG. 8 illustrates the positioning of the secondary container 12 to be centrifuged in a vertical position above the centrifuge 23, which is below the secondary container 12 in the proper position for transfer.

Because of the particular sequential arrangement of the layers in the secondary container 12, the platelet-rich plasma 17 and the adult mesenchymal autologous stem cells 16 are easily transferred. In addition, because the secondary container 12 is also preset to an evacuation level, the container only partially fills after blood collection. This allows the gas in the "head space" to remain significantly above zero during transfer when its volume is expanded, thereby allowing fast and complete transfer into the secondary container 12. This is dictated by the ideal gas law and the Poiseuille-Hagen equation.

Transfer of the contents or fragments of the secondary container (i.e. the platelet-rich plasma and adult mesenchymal autologous stem cells) continues into the collecting device with a housing and a member 20, wherein the member is moveable in a distal direction in order to transfer the medium until only the red blood cells remain in the secondary container 12. The transfer is performed in such a way that it also prevents accidental contamination by blood borne pathogens by prior use on or by another patient.

The transfer of the plasma fraction consisting of the platelet-rich plasma 17 and adult mesenchymal autologous stem cells 16 to the collecting device with a housing and a member 20 is complete, thereby allowing maximum yield and maintenance of the appropriate mediums. The collecting device 20 is then able to dispense its medium into an area that includes but is not limited to; musculoskeletal 24, ligament 25, tendon 26, cartilage 27, meniscus 28, disc 29(FIG.16), and organ of a living organism for regeneration of said area and/or; the container containing the medium of plasma, adult mesenchymal autologous stem cells and sodium chloride or similar medium can now be dispensed into a living organism via intravenous or intra-arterial (FIG. 17); or, the container containing the medium of plasma, adult mesenchymal autologous stem cells and albuterol or sodium chloride or similar medium can now be dispensed into a living organism via inhalation by way of nebulizer (FIG. 19); or, the container containing the medium of plasma is able to dispense its medium into a second medium housed in a container 30 (FIG. 13) for multiplying, regenerating, duplicating and incubating adult mesenchymal autologous stem cells for future use. The transfer occurs without venting, maintaining sterility and noncontamination of the sample.

The invention also provides a ready-to-use step-by-step system and methodology for deriving, preparing and administering adult mesenchymal autologous stem cells suitable for regenerating tissue in a living organism as shown in FIG. 20. The methodology comprises a primary container 10, the secondary container 12 and the transfer device 10.

In one embodiment, the system is packaged in such a way that the accoutrements include all the components required for deriving, preparing and administering adult mesenchymal autologous stem cells suitable for regenerating tissue in a living organism as shown in FIG. 20. Of course, the components can be arranged in a wide variety of manners.

In one aspect, the invention provides a system and methodology for deriving, preparing and administering platelet rich plasma and adult mesenchymal autologous stem cells suitable for regenerating tissue in a living organism.

Step 1 comprises collecting blood into the primary container 10 into the secondary container 12, followed by centrifugation to obtain platelet rich plasma and adult mesenchymal autologous stem cells. The components for the initiation of the process comprise an alcohol swab 34 to cleanse the venipuncture site, a multiple sample blood collection needle 10 as part of the primary container (21 gauge.times.1"), the secondary container 12 containing the anticoagulant (e.g. citrate), and a bandage 35 to cover the venipuncture site. The venipuncture site is cleansed with the sterile alcohol swab 34. The needle cover is removed from the end of the primary container. The needle at the end of the primary container 10 is then inserted into the patient's vein and the opposite end of the primary container 10 is inserted into the seal 13 of the secondary container 12. Blood then fills the secondary container12, after which the needle is withdrawn from the seal 13 of the secondary container and afterwards the needle is removed from the patient's vein and retracted into its holder. The vein is closed with the bandage 35. The secondary container 12 is

placed in a vertical position with its sealed opening 13 in the top most position as best shown in FIG. 8. This positions the secondary container 12 for the centrifuging of the secondary container's contents therein. FIG. 8 illustrates the positioning of the secondary container 12 to be centrifuged in a vertical position above the centrifuge 23, which is below the secondary container 12 in the proper position for transfer.

Because of the particular sequential arrangement of the layers in the secondary container 12, the platelet rich plasma 17 and the adult mesenchymal autologous stem cells 16 are easily transferred. In addition, because the secondary container 12 is also preset to an evacuation level, the container only partially fills after blood collection. This allows the gas in the "head space" to remain significantly above zero during transfer when its volume is expanded, thereby allowing fast and complete transfer into the secondary container 12. This is dictated by the ideal gas law and the Poiseuille-Hagen equation.

The container 12 is centrifuged at about 3000 xG for about 15 minutes and the platelet rich plasma and adult mesenchymal autologous stem cells are separated from the red blood cells.

The secondary container after being centrifuged contains residual gas 18, platelet rich plasma 17, buffy layer with adult mesecnhymal autologous stem cells 16 and red blood cells 15.

The components of the second step in the methodology for deriving, preparing and administering platelet rich plasma and adult mesenchymal autologous stem cells suitable for regenerating tissue in a living organism include a collecting device with a housing and a member 20 is complete, thereby allowing maximum yield and maintenance of the appropriate mediums.

Transfer of the contents or fragments of the secondary container (i.e. the platelet rich plasma and adult mesenchymal autologous stem cells) continues into the collecting device with a housing and a member 20, that has a the member that is moveable in a distal direction in order to transfer the medium until only the red blood cells remain in the secondary container 12. The transfer is performed in such a way that it also prevents accidental contamination by blood borne pathogens by prior use on or by another patient.

The transfer of the plasma fraction consisting of the platelet rich plasma 17 and adult mesenchymal autologous stem cells 16 to the collecting device with a housing and a member 20 is complete, thereby allowing maximum yield and maintenance of the appropriate mediums. The collecting device 20 is then able to dispense its medium into an area that includes but is not limited to; musculoskeletal 24, ligament 25, tendon 26, cartilage 27, meniscus 28, disc 29(FIG.16), and organ of a living organism for regeneration of said area and/or; the container containing the medium of platelet rich plasma, adult mesenchymal autologous stem cells and sodium chloride or similar medium can now be dispensed into a living organism via intravenous or intra-arterial (FIG. 17); or, the container containing the medium of plasma, adult

mesenchymal autologous stem cells and sodium chloride or similar medium can now be dispensed into a living organism via inhalation by way of nebulizer (FIG. 19); or, the container containing the medium of plasma is able to dispense its medium into a second medium housed in a container 30 (FIG. 13) for multiplying, regenerating, duplicating and incubating adult mesenchymal autologous stem cells for future use. The transfer occurs without venting, maintaining sterility and non-contamination of the sample.

In a second embodiment, the chamber 32 may also contain one or more of an antibiotic, an analgesic, a cancer therapeutic, a platelet-growth factor and a bone morphogenic protein. Other therapeutic agents which can be topically administered may also be included. Examples of antibiotics include, but are not limited to, ampicillin, erythromycin and tobramycin. Analgesics include, but are not limited to, aspirin and codeine. Cancer therapeutics include, but are not limited to, 5-fluor-uracile.

In operation, a patient's blood 11 is collected into the secondary collection device 12 by conventional venipuncture technique as described above. The anticoagulant in the primary collection device 12 thins the blood before centrifugation.

In that embodiment, the reservoir 32 may also contain one or more of an antibiotic, an analgesic, a cancer therapeutic, a platelet-growth factor and a bone morphogenic protein. Other therapeutic agents which can be topically administered may also be included. Examples of antibiotics include, but are not limited to, ampicillin, erythromycin and tobramycin. Analgesics include, but are not limited to, aspirin and codeine. Cancer therapeutics include, but are not limited to, 5-fluor-uracile.

I, Jimmy Lee Taylor, do hereby transfer & relinquish to my only heir & beneficiary, Zachary Sebastian Taylor, the following

- 1) All proprietary patents (listed on separate page) & intellectual property in my name, or that of my non-profit American Hospitals Incl. pertaining to stem cell IP, including, but not limited to, write pages and clinical studies on disease & treatment research, proprietary formulas and chemical solutions, and proprietary natural stem cell & hormone extraction techniques.
- 2) Specialized Power of Attorney to action my behalf, whether or not Lamincapacitated. In matters of, but not limited to, banking and purchasing
- Any liquid assets in bank accounts, foreign & domestic (listed on separate page).
- 4) Any and all proceeds from my non-profit American Hospitals Intl., which I would otherwise receive.

This agreement is lawfully binding and supersedes and overrides any previous such agreements.

Dr. Jon Taylor

4/30/18

ebastian Taylor - WITNESS

RECORDED: 12/04/2018